ADVANTAGES AND DISADVANTAGES OF PESTICIDE ANALYSIS METHODS USED IN AGRICULTURAL SAMPLES

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Abstract

Pesticides are substances (herbicides, fungicides, insecticides, plant growth regulators etc.) used primarily for pest control that can occur in both animals and plants. Unfortunately, besides beneficial effects, their use also has many disadvantages, these being toxic to humans and environment. For this reason, it is very important to have precise and accurate analytical methods for pesticide determination and quantification. The main purpose of this paper was to provide a description of the most commonly used methods of analysis and sample preparation for qualitative and quantitative determination of pesticides. The field of agriculture was analysed as a field of use of pesticides. Thus it has been highlighted that current analysis methods heavily rely on the use of gas chromatography analysis tools and in regard to the methods of agricultural samples preparation, these are in general extraction methods. The paper presents the advantages and disadvantages of the mentioned methods, in qualitative and quantitative assessment of pesticide

Key words: pesticides, agriculture, analytical method, chromatography.

INTRODUCTION

Pesticides are substances used primarily for pest control (US Environmental, 2007) that can occur in both animals and plants.

There are several classes of pesticides, the most common ones being:

- Organochlorine pesticides: DDT is the most used pesticide from this class and although its use has been restricted, there are countries that are thinking of reintroducing it (Turusov et al., 2002; Van den Berg, 2009);

- Organophosphorus pesticides: although they are said to be a more ecological option, many of the substances from this class have been associated with an endocrine disrupting potential (Mnif et al., 2011; Karami-Mohajeri et al., 2011);

- Carbamate pesticides: unfortunately the use of this class of pesticide can lead to a series of negative activity on the human body such as: possible reproductive disorders (Jamal et al., 2015), genotoxic effects in hamster ovarian cells (Soloneski et al., 2015) and last but not least increased risk for dementia (Lin et al., 2016).

There are three ways in which humans can be affected by pesticides (Yusà, Coscollà and Millet, 2014) and the most important source is through diet or ingestion.

Another way is through dermal contact, a way that it's more and more encountered due to household use of pesticides.

And a third way is by inhalation of contaminated air, particularly for those are staying nearby the agriculture areas.

Pesticides besides activities that have a negative impact on the health of people, affect on long term all type of water (surface and underground), air, soil and also soil organisms (Sarfraz et al., 2009)

To understand better the importance of pesticide use a short history of these compounds have been summarised in Table 1.

| Period | Type of pesticide used | | | |
|--------------------------|---|--|--|--|
| Ancient time | Ashes, common salts and bitters | | | |
| 1 st century | Arsenic, suggestion of soda and olive oil for treatment of legumes (Pliny the Elder, a Roman naturalist-Historia naturalis) | | | |
| 16 th century | Arsenicals and nicotine in the form of tobacco extracts (Chinese farmers) | | | |
| 1850 | Pyrethrum, soap and a wash of tobacco, sulfur and lime also used | | | |
| 1867 | The pigment Paris green (impure form of copper arsenite), Paris green and kerosene oi emulsion | | | |
| 1896 | Bordeaux mixture (CuSO ₄ and Ca(OH) ₂) and selective chemical herbicides | | | |
| 1900 | Dilute sulfuric acid, copper nitrates and potassium salts | | | |
| 1900-1950 | Sodium arsenite solutions become the standard herbicides and are used in large quantities | | | |
| 1913 | Organomercury seed dressing | | | |
| 1913-1939 | Dithiocarbamates fungicides used in US | | | |
| 1939 | Insecticidal potential of DDT discovered in Switzerland. Chlorinated hydrocarbons (DDT, BHC, dieldrin, aldrin and chlordane) | | | |
| 1950s | Fungicides captan, glyodin and organophosphorus insecticide: malathion | | | |
| 1961 | DDT registered for use on 34 different crops as pesticide usage dramatically increases | | | |
| 1962 | Bio accumulation and long-term toxicity and pest resistance became evident. Stoppa of DDT usage and other chlorinated compounds by farmers. Favor of the use of Organophosphates and Carbamates | | | |
| 1972 | Environmental Protection Agency revoked the use of DDT on all food sources in the United States. The World Health Organization, however, still reserves the right to us DDT on particularly virulent outbreaks of malaria | | | |
| 1972-1980 | Herbicidal sulfonylureas, neonicotinoids, glyphosate, synthetic fungicides such as metaxyl and triadimefron and light-stable pyrethroid pesticides are introduced | | | |
| 1990s | Integrated pest management, intensified research on biological pest control methods and other alternatives to pesticides | | | |
| 1990-1995 | Increased interest in Integrated Pest Management (IPM) programs | | | |
| 2000 | Wide spread usage of IPM techniques organic farming excluding the usage of synthet pesticides. | | | |
| 2010-2015 | Involvement of genetic engineering and biotechnological methods to control the usage of pesticides eg. baculoviruses | | | |

Table 1. Brief history of pesticide use (Jojiya et al., 2017)

EXTRACTION METHODS OF PESTICIDES

A novel method for sample extraction that it is used more and more is QuEChERS (Sherish et al., 2017).

One study use QuEChERS for the determination of seven pesticides from Okavango Delta water samples and present a detection limit situated between 0.102 µg/L-1.693 µg/L and a recovery value situated between 61% and 95% (V.C. Obuseng et al., 2013). Correia-Sa et al. have analysed soil samples with organic carbon over 2.3% and obtained 3.42-23.77 µg/kg limit detection and 70%-120% recovery (Correia-Sa et al., 2012).

Fresh peppermint samples were analysed by Magdalena Slowik-Borowiec et al. using this type of extraction method and obtain 0.01 mg/kg limit of detection and 100% recovery (Slowick-Borowiec et al., 2012). This method has several advantages such as: environmentally friendly and is simple and fast.

Another type known and used extraction method is Supercritical Fluid Extraction (SFE) which can be used for both solid and semi-solid samples. It is a recently developed method. The main advantage is that SFE is simple and less time consuming. Teresa Castelo-Grande et al. developed a SFE method for pesticides from soil and obtain a recovery of atrazine higher that 96% (Teresa Catselo-Grande et al., 2005). Tatsuo Yoshida et al. obtain a value for recovery of Isotianil extracted from rice and rice cultivation soil between 95.1% and 99.3% (T. Yoshida et al., 2013).

Hiroaki Chikushi et al. (H. Chikushi et al., 2009) evaluated the presence in water sample and proposed a method with low limit of detection: $0.002-2.3 \mu g/l$.

A quite new extraction method is accelerated solvent extraction (ASE). Important points in

this technique are temperature and pressure and also have advantages like speed and simplicity. Another plus is represented by the volume of reactive which is relatively low. Michelle L. Hladik et al. have analysed environmental sediment samples and obtain 81-101% recovery and 0.6-3.1 μ g/kg limit of detection (Michelle L. Hladik et al., 2012).

Beside the extraction methods mentioned above, there are presented in Table 2 other data obtained by using different types of extractions.

| Extraction method | Recovery/Limit of detection (LOD) | Sample | Advantages/Disadvantages | Reference |
|--|--|--|--|--------------------------------------|
| Headspace Single-Drop Micro-extraction (HS-SDME) | LOD = 0.07-12.54 µg/kg Recovery = 74-102% | Honey | Advantages: Possibility of using various solvents; Very good for extraction of diazinon; Possibility of extracting volatile and water-soluble analytes. | Amvrazi et al., 2012 |
| Solid Phase Extraction (SPE) | LOD = 0.01-0.088 µg/L Recovery = 74.2- 116.4% | Water samples | Advantages: Can be used to determine may types of pesticides; Present very good limit of detection; Rapid and efficient method. | Lopez- Mesas et al., 2007 |
| Dispersive liquid–liquid micro-extraction (DLLME) | LOD = 0.0032- 0.0174 µg/L Recovery = 84-108 % | Water samples | Advantages: Fast; Sensitive; Multi- residue method; Very good recovery. Disadvantages: Limited solvent choice; It's not suitable when the matrix composition is complex. | Abdullash et al., 2017 |
| Solvent-based de- emulsification dispersive liquid- liquid micro- extraction (SD- DLLME) | Recovery = 60-120% | Water samples | Advantages: Environmentally friendly; Less expensive than other techniques; Can be applied also for pharmaceuticals and personal care products. Disadvantages: same as DLLME. | Caldas et al., 2016 |
| Accelerated solvent extraction (ASE) | LOD = 0.6-3.1 µg/kg Recovery = 81-101% | Sedimentation of agricultural drainage samples | Advantage: Small volumes of solvents; Fast, easy and simple. Disadvantages: Relatively high price | Hladik et al., 2012 |
| | LOD = 0.8-3.4 µg/kg Recovery = 75-102% | Sediment samples from the estuary | of the equipment. | |
| Solid-phase extraction (D- SPE) by quick, easy, cheap, effective, rugged and safe (QuEChERS) | LOQ = 0.1-100 µg/kg | Rice paddy soils | Advantages: Alternative materials are more effective and less expensive than traditional sorbents Disadvantages: Limited solvent choice for extraction. | Arias et al., 2014 |
| QuEChERS | Recovery = 70-120% | Cereals (corn, wheat flour and rice) | Advantages: Multi-residue analysis; Simple and with satisfactory accuracy. | He et al., 2015 |
| Direct immersion solid-phase micro-extraction (DI-SPME) | LOD = 0.015-0.13 µg/L | Aqueous samples | Advantages: Can be applied on all types of water samples; Multi-residue analysis. Disadvantages: Relatively expensive because of fiber cost; Matrix effects. | Tankiewicz et al., 2013 |
| Microwave- assisted extraction (MAE) | Recovery = 81.5- 108.4% | Grass samples; Vegetation from the contaminated industrial area of Torneiros | Both methods are suitable for chlorinated pesticides analysis. Advantages MAE: shorter extraction times, higher extraction rates. | Barriada- Pereira et al., 2003 |
| Soxhlet extraction | Recovery = 75.5- 132.7% | | | |

Table 2. Extraction method of pesticide

DETERMINATION METHODS OF PESTICIDES

Javad Ghodsi and Amir Abbas Rafati have developed a method for the determination of diazinon made by use of a MWCNTs/TiO2NPs nanocomposite sensor (J. Gjodsi et al., 2017).

Comparative with other methods used such as gas chromatography, high-performance liquid chromatography (HPLC), mass spectrometry method, spectrophotometry, infrared spectroscopy and an enzyme immunoassay (M. Khadem et al., 2017; T.D. Lazarevic-Pasti et al., 2013; G. Erdogdu, 2003) that are expensive, time-consuming and with the need of trained employers, this method has shown to be sensitive, fast and use an easy fabricate sensor that is not so expensive. Also the method presents a good limit of detection of 3 nM. The real samples have included city piped and agricultural water.

Another determination method that uses HPLC/MS/MS was developed by Hwa-mi Lee et al. They analyse 56 residual pesticides from commercial crops and obtain a recovery value between 65-82% and a detection limit up to $11.54 \mu g/kg$ (Hwa-mi Lee et al., 2013).

Hirahara et al. described a screening method for determination of 200 pesticides using GC/MS/MS.

The method present a recovery value situated in 50-150% interval and a good limit of quantification of 0.01 mg/kg (Hirahara et al., 2006).

There are many methods for determination of pesticides, some of which are presented in the Table 3.

| Recovery/ Limit of detection (LOD) | Sample | Advantages/Disadvantages | Reference |
|--|--|--|---|
| | | | |
| LOD = 0.07-19 µg/kg | Honey | Very good for determination of organochlorines and organophosphorus pesticides; Highly sensitive; Low detection limit. | Amvrazi et al., 2012 |
| LOD = 0.6-3.1 µg/kg Recovery = 81-101% | Sedimentation of agricultural drainage samples | Very good recovery value; Sensitive method. | Hladik et al., 2012 |
| LOD = 0.8-3.4 µg/kg Recovery = 75-102% | Sediment samples from the estuary | | |
| LOD = 0.001-0.144 $\mu g/L$ Recovery = 83-101% | Environmental or drinking water samples | Advantages: Low limit of detection Disadvantages: Relatively high price of the equipment | Moreno- Gonzalez et al., 2012 |
| Recovery = 74-111% | Vegetable and fruit samples | Multi-residue method; suitable for routine quantitative analyses of pesticide | Sicaperumal et al., 2015 |
| LOD = 0.4-48.2 µg/kg Recovery = 70-110% | | Advantages: The method is repeatable; | |
| LOD = 1-115 µg/kg Recovery = 70-110% | Grape, lemon, onion and tomatoes | Can be used in many types of | Lesueur et al., 2008 |
| Recovery = $96 \pm 9\%$ | Cereal samples (wheat, rye, barley, oats, maze, buckwheat) | Improved analytical performance parameters Multi-residue method. | Walorczyk et al., 2012 |
| | LOD = 0.6-3.1 µg/kg Recovery = 81-101% LOD = 0.8-3.4 µg/kg Recovery = 75-102% LOD = 0.001-0.144 µg/L Recovery = 83-101% Recovery = 74-111% LOD = 0.4-48.2 µg/kg Recovery = 70-110% LOD = 1-115 µg/kg Recovery = 70-110% | LOD = $0.6-3.1 \ \mu g/kg$ Recovery = $81-101\%$ Sedimentation of agricultural drainage samplesLOD = $0.8-3.4 \ \mu g/kg$ Recovery = $75-102\%$ Sediment samplesLOD = $0.001-0.144$ $\ \mu g/L$ Recovery = $83-101\%$ Environmental or drinking water samplesLOD = $0.001-0.144$ $\ \mu g/L$ Recovery = $83-101\%$ Environmental or drinking water samplesRecovery = $74-111\%$ Vegetable and fruit samplesLOD = $0.4-48.2 \ \mu g/kg$ Recovery = $70-110\%$ Grape, lemon, onion and tomatoesLOD = $1-115 \ \mu g/kg$ Recovery = $70-110\%$ Cereal samplesRecovery = $96 \pm 9\%$ Cereal samples | LOD = 0.6-3.1 μ g/kg Recovery = 81-101%Sedimentation of agricultural drainage samplesVery good recovery value; Sensitive method.LOD = 0.8-3.4 μ g/kg Recovery = 75-102%Sediment samples from the estuaryVery good recovery value; Sensitive method.LOD = 0.001-0.144 μ g/L Recovery = 83-101%Sediment or drinking water samplesAdvantages: Low limit of detection Disadvantages: Relatively high price of the equipmentRecovery = 74-111%Vegetable and fruit samplesMulti-residue method; suitable for routine quantitative analyses of pesticideLOD = 0.4-48.2 μ g/kg Recovery = 70-110%Grape, lemon, onion and tomatoesMulti-residue method; suitable for routine quantitative analyses of matricesLOD = 1-115 μ g/kg Recovery = 70-110%Cereal samplesAdvantages: The method is repeatable; Can be used in many types of matricesRecovery = 96 ± 9%Cereal samples (wheat, rye, barley, oats, maze,Improved analytical performance parameters Multi-residue method |

Table 3. Determination method of pesticide

CONCLUSIONS

Due to the numerous negative effects on human and environmental, it is important and there is still a need to develop precise, sensible and robust extraction and analysis methods to determine the amount of pesticides and to keep them in conformity with applicable laws. Taking into account all these aspects, in this paper we have briefly discussed the most commonly used extraction and determination method for pesticides mainly from agriculture domain and also from other fields.

ACKNOWLEDGEMENTS

This work was carried out in Nucleu Programme TEX-PEL-2020, implemented with the support of MCI, project nr. PN 18 23 01 02, entitled "Exploitation of filamentous fungi for the production of bio-composite materials".



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