

## THE INFLUENCE OF COVER CROPS AND ANTAGONISTIC FUNGI ON THE HEALTHINESS OF CARROT (*DAUCUS CAROTA* L.)

Elżbieta PATKOWSKA, Elżbieta MIELNICZUK, Agnieszka JAMIOLKOWSKA

University of Life Sciences in Lublin, 7 K. St. Leszczyńskiego Street, Lublin, Poland

Corresponding author email: elzbieta.patkowska@up.lublin.pl

### Abstract

Cover crops and antagonistic fungi have a very positive effect on the soil environment. Such cover crops as oats, rye, common vetch, tansy phacelia, white mustard, sunflower and fodder radish can be used in the cultivation of different species of vegetables. They increase the biological activity of the soil by stimulating the growth and development of microorganisms antagonistic and can protect cultivated plants from soil-borne phytopathogens. The purpose of the field and laboratory studies was to determine the antagonistic activity of selected fungi species occurring in the soil under carrot cultivated with the use of oats, tansy phacelia and spring vetch as cover crops. The healthiness of the roots of this vegetable was also investigated. Cover crops contributed to the increase of the population of antagonistic fungi (*Trichoderma* spp., *Clonostachys* spp., *Myrothecium* spp. and *Penicillium* spp.) in the soil. Regardless of the experimental treatment, those antagonistic fungi were most effective in limiting the growth of *Sclerotinia sclerotiorum*, *Alternaria dauci* and *A. radicina*. The effect of those fungi was a little smaller towards *Fusarium oxysporum* and *Rhizoctonia solani*. Oats and spring vetch were most effective in limiting the occurrence of soil-borne fungi. Cover crops had a positive effect on the healthiness of carrot roots. *Alternaria dauci*, *A. alternata*, *A. radicina*, *Fusarium oxysporum*, *Globisporangium irregulare*, *Neocosmospora solani*, *Phytophthora* sp., *Rhizoctonia solani* and *Sclerotinia sclerotiorum* proved to be the most harmful towards the studied underground parts of carrot. Oats proved to be the most effective in inhibiting the occurrence of the pathogenic fungi for *Daucus carota* L.

**Key words:** *Daucus carota* L., cover crops, soil-borne fungi, phytopathogens, healthiness of plants.

### INTRODUCTION

Cover crops and antagonistic fungi have a very positive effect on the soil environment. Such cover crops as oats, rye, common vetch, tansy phacelia, white mustard, sunflower and fodder radish can be used in the cultivation of different species of vegetables. They increase the biological activity of the soil by stimulating the growth and development of microorganisms antagonistic and can protect cultivated plants from soil-borne phytopathogens (Hallama et al., 2019; Schmidt et al., 2018).

Cover crops can increase the number antagonistic microorganisms in the soil and improve the quality of the plants yield (Himmelstein et al., 2016; Oliveira et al., 2016).

### MATERIALS AND METHODS

The field experiment was conducted in district of Lublin (22°56'E, 51°23'N, Central Eastern Poland, 200 m a.s.l.), on Haplic Luvisol formed from silty medium loams. The object of the

studies was the soil taken from the field where carrot cv. 'Flakkee 2' was cultivated.

The experiment took into consideration cover crops such as oats (*Avena sativa* L.), spring vetch (*Vicia sativa* L.), tansy phacelia (*Phacelia tanacetifolia* Benth.) and one system of soil tillage, i.e.: tillage before winter (ploughing) and spring tillage (a combined cultivator). The conventional cultivation, i.e. without any cover crops, was the control.

Microbiological analysis of the soil was made according to the method described by Patkowska and Błazewicz-Woźniak (2014). All fungi isolates from the genera of *Clonostachys*, *Myrothecium*, *Penicillium* and *Trichoderma*, obtained from particular experimental treatments, were used to establish their antagonistic effect towards such fungi as *Alternaria dauci*, *Alternaria radicina*, *Fusarium oxysporum*, *Rhizoctonia solani* and *Sclerotinia sclerotiorum*. Moreover, the healthiness of carrot roots were determined.

The mycological analysis was conducted according to the method described by Patkowska and Krawiec (2016) for pea. This

analysis made it possible to determine the quantitative and qualitative composition of fungi infecting the underground organs of carrot.

## RESULTS AND DISCUSSIONS

Cover crops contributed to the increase of the population of antagonistic fungi (*Trichoderma* spp., *Clonostachys* spp., *Myrothecium* spp. and *Penicillium* spp.) in the soil. Regardless of the experimental treatment, those antagonistic fungi were most effective in limiting the growth of *Sclerotinia sclerotiorum* (Figure 1), *Alternaria dauci* and *A. radicina*. The effect of those fungi was a little smaller towards *Fusarium oxysporum* and *Rhizoctonia solani*. *Trichoderma* spp. and *Clonostachys* spp. can limit the occurrence of soil pathogens and they improve the plants' healthiness (Sarma et al., 2014; Smitha et al., 2014).

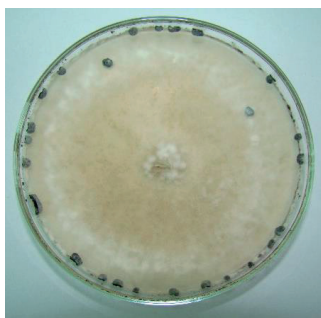


Figure 1. The 10-days colony of *Sclerotinia sclerotiorum* on the malt medium (photo by E. Patkowska)

Oats and spring vetch were most effective in limiting the occurrence of soil-borne fungi. A similar effect of the studied fungi antagonistic towards different species of pathogenic fungi was shown by Patkowska and Konopiński (2014) in the cultivation of scorzonera where cover crops were used. Banaay et al. (2012), Krauss et al. (2013) and Teshome et al. (2013) report that the ability of *Trichoderma* spp., *Clonostachys* spp. and *Penicillium* spp. to inhibit the growth and development of plant pathogens is based on antibiosis, competition and parasitism.

Cover crops had a positive effect on the healthiness of carrot roots. After the harvest of carrot, among fungi considered to be pathogenic, species from the genera of *Alternaria*,

*Fusarium*, *Phytophthora*, *Rhizoctonia* and *Sclerotinia* were isolated from diseased roots (Table 1). A little more the enumerated fungi were isolated from the infected roots of carrot (Figure 2) cultivated conventionally or after tansy phacelia as a cover crops, while the least after oats.



Figure 2. Sclerotia and mycelium of *Sclerotinia sclerotiorum* on the carrot roots (photo by E. Patkowska)

Table 1. Fungi isolated from diseased roots of carrot after harvest

Fungus species	Experimental treatment/Number of isolates										Total (%)
	Oats		Spring vetch		Tansy phacelia		Control		Total		
	a*	b	a	b	a	b	a	b	a	b	
<i>Acremonium murorum</i> (Corda) W. Gams	-	-	-	-	3	-	4	2	8	2	10 (1.5)
<i>Alternaria alternata</i> (Fr.) Keissl.	1	-	3	-	5	2	16	6	25	8	33 (4.8)
<i>Alternaria chartarum</i> Preuss	-	-	-	-	3	-	9	3	12	3	15 (2.2)
<i>Alternaria dauci</i> (J.G. Kühn) J.W. Groves & Skolko	1	-	1	-	4	2	11	6	17	8	25 (3.6)
<i>Alternaria radicina</i> Meier, Drechsler & E.D. Eddy	1	-	1	-	6	4	17	14	25	18	43 (6.3)
<i>Cladosporium cladosporioides</i> (Fresen.) G.A.de Vries	-	-	-	2	2	3	6	3	8	8	16 (2.3)
<i>Clonostachys rosea</i> (Link)	11	9	9	7	6	3	-	-	26	19	45 (6.6)

Schroers, Samuels, Seifert & W. Gams														
<i>Cylindrocarpum didymum</i> (Harting) Wollenw.	-	-	-	-	3	2	8	4	11	7	18	(2.6)		
<i>Epicoceum nigrum</i> Link	-	-	1	-	2	2	7	3	10	5	15	(2.2)		
<i>Fusarium avenaceum</i> (Fr.) Sacc.	-	-	-	-	2	2	6	4	8	6	14	(2.0)		
<i>Fusarium oxysporum</i> Schltdl.	4	2	6	4	8	4	10	6	28	16	44	(6.4)		
<i>Neocosmospora solani</i> (Mart.) L. Lombard Crous	-	-	2	2	3	5	8	3	13	10	23	(3.3)		
<i>Penicillium aurantiogriseum</i> Dierckx	2	1	3	1	4	2	8	4	17	8	25	(3.6)		
<i>Penicillium chrysogenum</i> Thom	-	-	-	1	2	1	5	3	7	5	12	(1.7)		
<i>Penicillium meleagrinum</i> Biourge	-	-	-	-	3	2	9	5	12	7	19	(2.8)		
<i>Penicillium simplicissimum</i> (Oudem.) Thom	-	-	-	-	1	-	6	4	7	4	11	(1.6)		
<i>Phytophthora</i> sp.	2	-	2	1	4	2	6	5	16	8	24	(3.4)		
<i>Rhizoctonia solani</i> J.G. Kühn	5	3	7	4	8	6	12	10	32	23	55	(8.0)		
<i>Rhizopus stolonifer</i> (Ehrenb.) Vuill.	2	3	3	2	8	3	16	8	29	16	45	(6.5)		
<i>Sclerotinia sclerotiorum</i> (Lib.) de Bary	2	2	5	3	7	5	14	12	28	22	50	(7.3)		
<i>Trichoderma harzianum</i> Rifai	7	5	6	4	5	3	2	-	20	12	32	(4.6)		
<i>Trichoderma koningii</i> Oudem.	14	12	12	10	10	9	3	3	39	34	73	(10.6)		
<i>Trichoderma viride</i> Pers.	10	7	8	5	6	4	2	-	26	16	42	(6.1)		
Total	62	44	69	46	105	66	186	108	422	264	686	(100.0)		
Total	106	115	171	294	686									

a\* - root; b - head of root

Studies by Koike et al. (2017) and Zafar et al. (2017) also pointed to considerable harmfulness of these fungi towards carrot plants. According to Rogers and Stevenson (2010), Tülek and Dolar (2015) and Zafar et al. (2017), fungi, especially *Alternaria dauci*, *A. radicina*, attacked carrots at all stages, causing damping-off, and rotting of roots, crowns, seedlings, petioles, leaves and crowns of maturing carrots.

Studies conducted by Patkowska et al. (2016) and Dawadi et al. (2019) showed that cover plants (rye, oats, white mustard) significantly reduce the population of pathogenic fungi in the soil environment, thus positively affecting the healthiness of the cultivated plants.

## CONCLUSIONS

The present studies confirmed the positive effect of cover crops on the growth and healthiness of *Daucus carota*.

Oats, tansy phacelia and spring vetch inhibited the occurrence and development of soil-borne fungi and - consequently - improved the healthiness of the examined plant.

## ACKNOWLEDGEMENTS

The studies were partially financed by the Polish Ministry of Science and Higher Education of Poland within grant No. NN 310 210 837 and statutory funds (OKF/DS/2) of the Department of Plant Pathology and Mycology, University of Life Science in Lublin, Poland.

## REFERENCES

- Banaay, C. G. B., Cuevas, V. C., Vera Cruz, M. C. (2012). *Trichoderma ghanense* promotes plant growth and controls disease caused by *Pythium arrhenomanes* in seedlings of aerobic rice variety apo. *The Philippine Agricultural Scientist*, 95, 175-184.
- Dawadi, S., Baysal-Gurel, F., Adesso, K. M., Oliver, J. B., Simmons, T. (2019). Impact of cover crop usage on soilborne diseases in field nursery production. *Agronomy* 2019(9), 753.
- Hallama, M., Pekrun, C., Lambert, H., Kandeke, E. (2019). Hidden miners – the roles of cover crops and soil microorganisms in phosphorus cycling through agroecosystems. *Plant Soil*, 434(1-2), 7.
- Himmelstein, J., Maul, J. E., Balci, Y., Everts, K. L. (2016). Factors associated with leguminous green

- manure incorporation and Fusarium wilt suppression in watermelon. *Plant Diseases*, 100, 1910-1920.
- Koike, S. T., Smith, R. F., Cahn, M. D., Pryor, B. M. (2017). Association of the carrot pathogen *Alternaria dauci* with new diseases, *Alternaria* leaf speck, of lettuce and celery in California. *Plant Health Progress*, 18(2), 136-143.
- Krauss, U., Hoopen, M., Rees, R., Stirrup, T., Argyle, T., George, A., Arroyo, C., Corrales, E., Casanoves, F. (2013). Mycoparasitism by *Clonostachys byssicola* and *Clonostachys rosea* on *Trichoderma* spp. from cocoa (*Theobroma cacao*) and implication for the design of mixed biocontrol agents. *Biological Control*, 67, 317-327.
- Oliveira, P., Nascente, A. S., Brito Ferreira, E. P., Kluthouski, J., Lobo jr., M. (2016). Response of soil fungi and biological processes to crop residues in no-tillage system. *Pesquisa Agropecuária Tropical*, 46(1), 57-64. (ww.agro.ufg.br/pat.).
- Patkowska, E., Błażewicz-Woźniak, M. (2014). The microorganisms communities in the soil under the cultivation of carrot (*Daucus carota* L.). *Acta Scientiarum Polonorum Hortorum Cultus*, 13(1), 103-115.
- Patkowska, E., Błażewicz-Woźniak, M., Konopiński, M., Wach, D. (2016). Effect of cover crops on the fungal and bacterial communities in the soil under carrot cultivation. *Plant, Soil and Environment*, 62(5), 237-242.
- Patkowska, E., Konopiński, M. (2014). Occurrence of antagonistic fungi in the soil after cover crops cultivation. *Plant, Soil and Environment*, 60(5), 204-209.
- Patkowska, E., Krawiec, M. (2016). Yielding and healthiness of pea cv. 'Sześciotygodniowy TOR' after applying biotechnical preparations. *Acta Scientiarum Polonorum Hortorum Cultus*, 15(2), 143-156.
- Rogers, P. M., Stevenson, W. R. (2010). Aggressiveness and fungicide sensitivity of *Alternaria dauci* from cultivated carrot. *Plant Diseases*, 94(4), 405-412.
- Sarma, B. K., Yadav, S. K., Patel, J. S., Singh, N. B. (2014). Molecular mechanisms of interactions of *Trichoderma* with other fungal species. *The Open Mycological Journal*, 8, 140-147.
- Schmidt, R., Gravuer, K., Bossange, A. V., Mitchell, J., Scow, K. (2018). Long-term use of cover crops and no-till shift soil microbial community life strategies in agricultural soil. *PLoS ONE*, 13(2), e0192953. (<https://doi.org/10.1371/journal.pone.0192953>).
- Smitha, C., Finosh, G. T., Rajesh, R., Abraham, P. K. (2014). Induction of hydrolytic enzymes of phytopathogenic fungi in response to *Trichoderma viride* influence biocontrol activity. *International Journal of Current Microbiology and Applied Sciences*, 3(9), 1207-1217.
- Teshome, E., Fininsa, Ch., Sahile, S. (2013). *In vitro* antagonistic potential of fungal isolates against *Botrytis fabae* Sard. *Asian Journal of Plant Pathology*, 7, 42-53.
- Tülek, S., Dolar, F. S. (2015). Detection and identification of *Alternaria* species causing diseases of carrot in Ankara province, Turkey. *Scientific Papers, Ser. B, Horticulture*, LIX, 263-268.
- Zafar, M. M., Abrar, M., Umar, M., Bahoo, M. A., Khan, N. A., Salahuddin, M., Bilal, A., Abdullah. (2017). Screening of different carrot varieties against *Alternaria* leaf blight and its chemical management. *Researcher*, 9(12), 8-14.