

## IDENTIFICATION OF ALTERNATIVE MEASURES FOR THE MANAGEMENT OF ROOT-KNOT NEMATODES ON SOLANACEOUS VEGETABLE CROPS IN SOUTHWEST BULGARIA

Lilyana KOLEVA\*, Georgi DIMITROV, Zhelyu AVRAMOV

University of Forestry, Faculty of Agronomy, 10 Kliment Ohridski, 1797, Sofia, Bulgaria

\*Corresponding author email: lmarkova@ltu.bg

### Abstract

*Vegetables from the Solanaceae family (eggplant, tomato, and potato) are among the crops in Europe which in terms of production rank first, and in Bulgaria their production is concentrated in the south-western part. The aim of the study was to identify alternative root-knot nematode control measures applicable in integrated pest management to improve plant health and reduce dependence on chemical pesticides in Solanaceae vegetable production. Based on the data of new research, a summary list of specific combinations of vegetable crops/species of root-knot nematodes in Bulgaria has been compiled. After analysing the problems and according to the innovative practices, methods to control a given root-knot nematode species in a certain crop were indicated. The studies enabled an inventory of potential alternative measures for integrated management and the creation of a dataset that will allow for the improvement of plant health and the reduction of dependence on chemical pesticides in the production of Solanaceae vegetables.*

**Key words:** alternative measures, root-knot nematodes, Solanaceae vegetable.

### INTRODUCTION

The economic damage caused by plant parasitic nematodes worldwide is estimated at 12.3% (Hassan et al., 2013; Singh et al. 2015). In heavily infected areas, yield losses of more than 50% are observed, in some cases the crop can be destroyed (Nicol et al., 2011). In addition, the presence of nematodes limits the cultivation of several crops on infected areas. Root-knot nematodes (*Meloidogyne* spp.) are distributed throughout the world and belong to the phytonematodes of greatest economic importance, followed by representatives of the genus *Pratylenchus* and the genus *Heterodera* (Ravichandra, 2014). Nematodes of the genus *Meloidogyne* Goldi 1877 are pests of economic importance for many crops grown in greenhouses and fields (Samaliev and Stoyanov, 2008; Mesa-Valle, et al. 2020). They are obligate, sedentary root parasites, polyphages. The most common species are *M. incognita*, *M. arenaria*, *M. javanica* and *M. hapla*, which develop several generations during one growing season and are characterized by high population density and dynamics. The host range of these species includes over 2000 plant species, and species from the Solanaceae family can be defined as the main hosts. Many of the world's

economically important agricultural crops belong to the family Solanaceae (*Solanum*, *Capsicum* and *Nicotiana*) with about 28 million hectares under cultivation worldwide (Motti, 2021). Vegetables from the Solanaceae family are among the crops in Europe that rank first in terms of production and production area (Santamaria & Signore, 2021). In recent decades, the consumption of fruits and vegetables has increased, with increased demand for tomato, pepper, and eggplant belonging to the family Solanaceae (Motti, 2021). However, losses are a major problem at all stages from production to consumption. Current chemical nematicides are insufficiently effective and must be optimized for specific pests and crops. The requirements to produce fresh vegetables without residual pollutants increase the interest of growers in the problems related to control measures and managing the impact of biotic stress factors. New and expanding trends in agriculture require research to provide cost-effective and easy-to-use alternatives to conventional synthetic pesticides or to identify measures compatible with integrated programs to protect and minimize the application of chemical agents. Information in the literature regarding the influence of alternative control measures on the development

of vegetable crops to overcome nematode damage is insufficient, there is a lack of specific data on vegetable production in Bulgaria.

The aim of the study was to identify alternative root-knot nematode control measures applicable in integrated pest management to improve plant health and reduce dependence on chemical pesticides in Solanaceae vegetable production.

## MATERIALS AND METHODS

The results of the present work were obtained based on the analysis of the scientific literature on the considered problem, the grouping of the obtained data, the monitoring of established root-knot nematode/crop combinations and laboratory analysis of soil and plant samples to determine the species composition of nematodes. The route studies and observations were conducted in the period 2021-2022. In the regions of Sofia, Pernik, Kyustendil, Samokov, Blagoevgrad, Gotse Delchev and Pazardzhik, 22 observation points were selected, which were planted by tomatoes and potatoes during the growing season. According to preliminary information and data of the Bulgarian Agency for Food Safety, root-knot nematodes were found on some of these areas. Investigations of the presence and distribution of root-knot nematodes of genus *Meloidogyne* included greenhouses and field areas of tomatoes, aubergines and potatoes intended for consumption and processing. The leading factor in choosing the plots with potatoes was that they should be grown on agricultural plots where monoculture tobacco was grown in the recent past. Figure 1 shows the surveyed regions.



Figure 1. Regions in Southwest Bulgaria that were subject to root-knot nematode monitoring

## RESULTS AND DISCUSSIONS

The following species of root-knot nematodes have been established in Bulgaria: southern root-nematode *Meloidogyne incognita* (Kofoid & White, 1919), peanut root knot nematode *Meloidogyne arenaria* (Neal, 1889), northern root-knot nematode *Meloidogyne hapla* Chitwood, 1949, *Meloidogyne javanica* Treub., 1885, Chitwood, 1949, thames' root-knot nematode *Meloidogyne thamesi* Chitwood, 1952. The cereal root-knot nematode *Meloidogyne naasi* Franklin, 1965 has not been established, but there are conditions for its development.

In Bulgaria in 1925, Chorbadjiev made the first reports of root-knot nematode damage to tobacco seedlings in the region of Shumen District. Later in 1940 Kovachevski found it on cucumbers in the village of Varbitsa, Gorna Oryahovitsa. Trifonova and Gospodinov (1955), Stoyanov (1962), Choleva (1973) and others reported intensive research on the bioecology of the species of the genus *Meloidogyne* (Mateeva, 2004).

The results for the identification of model combinations of vegetable crop/nematode species for the different regions of Bulgaria were presented in a summary list. The data from modern research on the species composition are reflected in Table 1.

The results of our tests confirmed the presence of nematodes of the genus *Meloidogyne* in the studied samples, which confirms their wide distribution of these pests in the region of Southwest Bulgaria. The species *Meloidogyne arenaria*, *Meloidogyne incognita* and *Meloidogyne javanica* were identified. Mixed infection (*M. arenaria* + *M. incognita*) was observed from the greenhouse samples. In the field samples, the dominant species was *M. arenaria*.

The review done here aims to evaluation the latest studies to alternative methods of root-knot nematode control. Table 2 lists those that are easily applicable and effective.

Table 1. Summary list of specific combinations vegetable crop/species of root-knot nematode in Bulgaria

Area	Species of root-knot nematode	Crop	Reference sources
Kresna	<i>Meloidogyne</i> spp.,	tomato	Markova L. et al. (2014)
Southern Bulgaria	<i>Meloidogyne</i> spp.,	tomato	Trifonova & Vulkova (2007); Voulkova & Trifonova (2009); Tringovska et al. (2015)
	<i>M. arenaria</i>	tomato	Choleva et al. (2005); Yankova et al. (2006); Baycheva et al. (2018)
	<i>Meloidogyne</i> spp.,	greenhouse vegetable crops	Choleva et al. (2004)
Southern Bulgaria	<i>Meloidogyne</i> spp.,	greenhouse vegetable crops	Samaliev (2009b)
	<i>M. hapla</i>	potatoes, tomatoes	Trifonova & Voulkova (2008); Markova & Samaliev (2011)
	<i>M. javanica</i>	aubergine	Mohamedova et al. (2016)
	<i>M. incognita</i>	cucumbers, tomatoes	Trifonova & Voulkova (2008); Trifonova & Vachev (2010); Panayotova et al. (2016)
	<i>M. incognita</i> , <i>M. arenaria</i> , <i>M. javanica</i> , <i>M. hapla</i> и <i>M. thamesi</i>	greenhouse vegetable crops and arable crops	Samaliev et al. (2018)
	<i>M. arenaria</i> , <i>M. incognita</i> , <i>M. javanica</i> and <i>M. hapla</i> , races 1 and 2 of <i>M. incognita</i> and race 1 and 2 of <i>M. arenaria</i>	greenhouse vegetable crops	Samaliev (2009a).
Plovdiv, Troyan and Samokov	<i>M. hapla</i> , <i>M. arenaria</i> and <i>M. incognita</i>	potato	Samaliev & Baicheva (2010); Samaliev & Kalinova (2013)
	<i>M. incognita</i> , <i>M. arenaria</i> and <i>M. hapla</i>	over 30 host plants	Stoyanov (1980).
	<i>M. arenaria</i>	cucumbers	Yankova, et al. (2014).

Table 2. Alternative methods for control of root-knot nematodes (*Meloidogyne* spp.) in major vegetable crops

Alternative control measure	Species of root-knot nematode	Crop	Reference sources
cultural practices	<i>Meloidogyne</i> spp.	host plants	Azlay et al. (2022)
solarization	<i>Meloidogyne</i> spp.	tomato	Yücel et al. (2007); Samaliev (2009b).
resistant varieties	<i>Meloidogyne</i> spp., <i>M. incognita</i>	tomato	Lizardo et al. (2022); Trifonova & Voulkova (2008); Trifonova & Vulkova (2007); Yankova et al. (2006)
sanitation, heat-based methods	<i>Meloidogyne</i> spp.	vegetable crops	Collange et al. (2011)
plant extracts and essential oils	<i>M. incognita</i> ; <i>M. incognita</i> race 2, <i>M. hapla</i>	tomato, cucumber,	Abo-Elyousr et al. (2009); Abo-Elyousr et al. (2010); Adegbite (2011); Azlay et al. (2022); Mostafa et al. (2017); Salim et al. (2016); Taniwiryono et al. (2009); Taye et al. (2012); Trifonova & Atanasov (2009); Trifonova (2012)

Alternative control measure	Species of root-knot nematode	Crop	Reference sources
aqueous extracts of garlic and <i>Ricinus</i> seeds	<i>M. incognita</i>	tomato	El-Nagdi & Youssef (2013)
organic amendment	<i>M. incognita</i>	tomato	Asif et al. (2016); Zakaria et al. (2013)
biological, chitinous material, seashell meal	<i>M. incognita</i>	tomato	Ladner et al. (2008)
biological management/ nematophagous bacteria and fungi	<i>Meloidogyne</i> spp.	host plants	Azlay et al. (2022); Moreira et al. (2015); Singh et al. (2019); Zakaria et al. (2013)
bioproducts of microbial origin	<i>M. incognita</i>	tomatoes	Radwan et al. (2012)
commercial products contain bacteria <i>Bacillus firmus</i> and <i>Pasteuria penetrans</i> , and <i>Purpureocillium lilacinus</i> mushroom	<i>Meloidogyne</i> spp.	host plants	Lamovšek et al. (2013); Samaliev, H. (1997); Samaliev & Baycheva (2004).
<i>Bacillus thuringiensis</i> /crystal protein/	<i>Meloidogyne</i> spp.; <i>M. arenaria</i>	tomato	Li et al. (2007); Mohamedova (2009)
rhizobacteria <i>Bacillus subtilis</i> <i>Bacillus altitudinis</i> AMCC1040 <i>Pseudomonas oryzae</i> and <i>Xenorhabdus nematophilus</i>	<i>M. javanica</i> <i>M. javanica</i>	aubergine host plants	Mohamedova et al. (2016) Samaliev et al. (2000)
Fungi antagonists, <i>Acremonium strictum</i> & <i>Trichoderma harzianum</i>	<i>Meloidogyne</i> spp., <i>M. incognita</i>	tomato	Goswami et al. (2008); Jindapunnapat et al. (2013); Singh et al. (2019); Trifonova & Vachev (2010)
Nematophagous fungi, <i>Pochonia chlamydosporia</i>	<i>Meloidogyne</i> spp., <i>M. arenaria</i> , <i>M. incognita</i>	organic vegetable production	Atkins et al. (2003); Sosnowska (2007); Trifonova (2014)
stimulating plant growth; rhizobacteria strains	<i>M. incognita</i>	tomato	Cetintas et al. (2018)
grafting root stock / <i>Solanum sisymbriifolium</i> , cucurbitaceous rootstocks	<i>M. incognita</i> , <i>Meloidogyne</i> spp.	tomato	Baidya et al. (2017)
cover and biofumigant crops	<i>Meloidogyne</i> spp.	tomato and potato	Daneel et al. (2018)
cover crops and green manure crops of the genus Brassica; companion plants	<i>Meloidogyne incognita</i> , <i>Meloidogyne javanica</i>	vegetable plasticulture	Monfort et al. (2007); Stirling & Stirling (2003); Tringovska et al. (2015)
antagonistic plants (sorghum, crotalaria, mucuna, guandu bean and neem)	<i>M. javanica</i> , <i>M. enterolobii</i> , <i>Meloidogyne</i> spp., <i>M. arenaria</i>	host plants,	Moreira et al. (2015); Yasmin et al. (2003); Al Body & Mateeva (2007).
<i>African marigold</i> , <i>Tagetes erecta</i>	<i>M. incognita</i> , <i>M. arenaria</i>	tomato, greenhouse vegetable	Natarajan et al. (2006); Al Body & Mateeva (2007)

Alternative control measure	Species of root-knot nematode	Crop	Reference sources
<i>Tagetes minuta</i> , <i>Datura metel</i> , <i>D. stramonium</i> and <i>Ricinus communis</i>	<i>Meloidogyne</i> spp.	tomato	Oduor-Owino (2003)
abamectin	<i>Meloidogyne</i> spp.	tomato	Qiao et al. (2012);
<i>Chlorella vulgaris</i> /algae/ and potassium humate	<i>M. arenaria</i> , <i>M. incognita</i> , <i>M. hapla</i>	greenhouse tomato	Choleva et al. (2007); Choleva et al. (2004)
fertilization, compost, fertilizers, biostimulators poultry manure	<i>Meloidogyne</i> spp., <i>M. javanica</i> , <i>M. incognita</i>	greenhouse tomato	Saeedzadeh et al. (2020); Markova, L. et al. (2014)
silver nanoparticles	<i>Meloidogyne</i> spp.	tomato	Chindo & Khan (1990)
selenium	<i>M. arenaria</i>	tomato	Bernard Monfort et al. (2019); Baycheva et al. (2018)

After analysis of literature data, we found large variations between studies. Many practices were listed as only partially effective; therefore, combining control methods in integrated nematode management is a challenge that requires a systematic approach and identification of key future research. There is growing concern among vegetable growers as registered chemical nematicides decline. Alternative methods, means and techniques based on innovative practices are needed to solve the problem.

Despite the availability of effective alternative methods for root-knot nematode control, limited progress has been made in their implementation. In part, this may be due to the fact that the plant parasitic nematodes control has traditionally been carried out primarily by chemical means. The trend worldwide for a chemical pesticide-free agricultural production and environment minimizes the use of pesticides (Lykogianni et al., 2021). The application of crop rotation is economically unprofitable for farmers, and the main part of greenhouse vegetables in Bulgaria is grown as a monoculture. The breeding of resistant varieties against these enemies is a difficult task (Samaliev & Stoyanov, 2008). This necessitates the search for alternative methods of control of *Meloidogyne* spp. In this aspect, among the non-chemical control measures is the biological method, especially in the context of integrated plant management. One of the many options for biological control is to specifically obtain and support or colonize biological active agents that suppress the development of plant parasitic nematodes. In this regard, the availability of different types of biological

agents with potential for successful control of these enemies is of interest. Nematode antagonists can be various types of bacteria, fungi, viruses, rickettsia, predatory nematodes such as mites, collembolans (Volpiano et al., 2019). Trifonova et al. (2009) conducted mycological studies in the southern regions of Bulgaria and found that *Fusarium oxysporum*, *Verticillium chlamydosporium* and *Gliocladium roseum* parasitized the eggs of female individuals of the genus *Meloidogyne*, with 7.6% to 23.5% of the eggs in subsequent generations died. Of the previously known microorganisms with an antagonistic effect against *Meloidogyne*, spp. non-pathogenic bacterial strains have increasingly been used in recent years (Sidhu, 2018). The use of rhizobacteria to biologically control plant diseases and to stimulate plant growth has been practiced with great success (Shaikh & Sayyed, 2015; Verma, 2019). The search for suitable rhizobacteria against plant-pathogenic nematodes started about 30 years ago (Zavaleta-Meija & VanGundy, 1982; Mohamedova & Samaliev, 2011; Sidhu, 2018), mainly the genera *Pseudomonas* spp. and *Bacillus* spp. They have properties that determine their success in practice - they inhibit egg hatching or produce metabolites toxic to nematodes but have no negative effects on soil and on plant growth and development. Studies carried out by Ahmed (1999) on the effect of the rhizobacterium *Bacillus subtilis* on damage caused by *Meloidogyne* spp. in tomato showed that even when the root-knot nematode population increased and damage was important, plant growth was enhanced (induced tolerance). There

is data in the literature that suggests that treatment of plants with synthetic phytohormones and organomineral fertilizers also induced resistance to pests including nematodes (Guimarães, 2010; Bhattacharya, 2021; Dar et al., 2021).

In greenhouse experiments, the influence of using the following fertilizers Kendal, 18 Biopower, Nutriphite, Hortiplus MIQL 2826, Max Fitus as inducers of resistance to *Meloidogyne incognita* was studied. The authors reported proven differences in nematode population numbers between the control and variants treated with Biopower and Nutriphite fertilizers (Assunção et al., 2010).

In Bulgaria, Markova et al. (2014) studied the effect of the liquid root biostimulator Fertiactyl® GZ on damage by root-knot nematodes *Meloidogyne* spp. in tomato plants grown in greenhouse conditions.

The authors found that the liquid root biostimulator had a stimulatory effect on the growth and development of plants infected with *Meloidogyne* spp., and the tested tomato cultivars Raleigh and Matthias after treatment with Fertiactyl® GZ showed no symptoms of damage.

The cultivation of resistant varieties as well as the observance of crop rotation are methods that limit the use of the chemical control method. The protection of vegetable crops of the Solanaceae family against plant parasitic nematodes should ensure the health and potential of the crops through the extensive implementation of alternative control measures such as sanitary measures, phytosanitary monitoring, crop rotation, mixed cropping, breeding of resistant or tolerant varieties, application of biological agents and other innovative practices. The emergence of nematode resistance to nematicides, the negative impacts on human health and the environment, and the drastic decrease in the availability of existing and new chemical pesticides in Europe lead to an increased need for alternative control options. In addition, there are legal requirements to regulate agricultural production according to IPM principles and to market plant products with little or no pesticide residues (Directive 2009/128/EC).

## CONCLUSIONS

The analysis of the species composition of root-knot nematodes in solanaceous vegetable crops in Southwest Bulgaria showed that the distribution and the population structure were relatively constant over time.

Limited progress has been observed in implementing alternative measures to control root-knot nematodes.

A range of data was created to enable the inclusion of alternative measures in integrated pest management (IPM) programs and to reduce reliance on chemical control.

## ACKNOWLEDGEMENTS

This study was conducted within the project NIS-B-1210 “Identification of alternative measures for the management of root-knot nematodes on solanaceous vegetable crops to reduce the use of chemical pesticides” funded by the Scientific Sector of the University of Forestry.

## REFERENCES

- Abo-Elyousr, K. A., Awad, M.E.M., & Gaid, M. A. (2009). Management of tomato root-knot nematode *Meloidogyne incognita* by plant extracts and essential oils. *The Plant Pathology Journal*, 25(2), 189-192.
- Abo-Elyousr, K.A., Khan, Z., & Abdel-Moneim, M.F. (2010). Evaluation of plant extracts and *Pseudomonas* for control of root-knot nematode, *Meloidogyne incognita* on tomato. *Nematropica*, 289-299.
- Adegbite, A.A. (2011). Effects of some indigenous plant extracts as inhibitors of egg hatch in root-knot nematode (*Meloidogyne incognita* race 2). *American Journal of Experimental Agriculture*, 1(3), 96-100.
- Ahmed, S. (1999) Wirkungen des Rhizobakteriums *Bacillus subtilis* auf den Befall von Tomatenpflanzen durch Wurzelgallen- (*Meloidogyne* spp.) und Wurzelläsions-Nematoden (*Pratylenchus* spp.) DISSERTATION, Humboldt-Universität zu Berlin, 169.
- Al Body, M., & Mateeva, A. (2007). Influence of water extracts from *Tagetes* and *Calendula officinalis* against *Meloidogyne arenaria*. *Scientific Works, Agricultural University, Plovdiv* (Bulgaria), 52, 203-208.
- Asif, M., Khan, A., Tariq, M., & Siddiqui, M. A. (2016). Sustainable management of root knot nematode *Meloidogyne incognita* through organic amendment on *Solanum lycopersicum* L. *Asian Journal of biology*, 1(1), 1-8.

- Assunção, A., L.C. Santos, M.R. Da Rocha, A.J.S. Reis, R.A. Teixeira & F.S.O. Lima., 2010. Efeito de Indutores de Resistência sobre *Meloidogyne incognita* em Canade-açúcar (*Saccharum* spp.). *Nematologia Brasileira*, Brasília, 34(1), 56-62.
- Atkins, S. D., Hidalgo-Diaz, L., Kalisz, H., Mauchline, T. H., Hirsch, P. R., & Kerry, B. R. (2003). Development of a new management strategy for the control of root-knot nematodes (*Meloidogyne* spp) in organic vegetable production. *Pest Management Science: Formerly Pesticide Science*, 59(2), 183-189.
- Azlay, L., El Boukhari, M. E. M., Mayad, E. H., & Barakate, M. (2022). Biological management of root-knot nematodes (*Meloidogyne* spp.): a review. *Organic Agriculture*, 1-19.
- Baidya, S., Timila, R. D., KC, R. B., Manandhar, H. K., & Manandhar, C. (2017). Management of root knot nematode on tomato through grafting root stock of *Solanum sisymbriifolium*. *Journal of Nepal Agricultural Research Council*, 3, 27-31.
- Baycheva, O., Samaliev, H., Udalova, Z., Trayanov, K., Zinovieva, S., & Folman, G. (2018). Selenium and its effect on plant-parasite system *Meloidogyne arenaria*-Tiny Tim tomatoes. *Bulgarian Journal of Agricultural Science*, 24(2), 252-258.
- Bernard, G. C., Fitch, J., Min, B., Shahi, N., Egnin, M., Ritte, I., ... & Bonsi, C. (2019). Potential nematicidal activity of silver nanoparticles against the root-knot nematode (*Meloidogyne incognita*). *Online Journal of Complementary & Alternative Medicine*, 2, 000531.
- Bhattacharya, A., & Bhattacharya, A. (2021). Role of plant growth hormones during soil water deficit: A review. *Soil Water Deficit and Physiological Issues in Plants*, 489-583.
- Cetintas, R., Kusek, M., & Fateh, S. A. (2018). Effect of some plant growth-promoting rhizobacteria strains on root-knot nematode, *Meloidogyne incognita*, on tomatoes. *Egyptian Journal of Biological Pest Control*, 28(1), 1-5.
- Chindo, P. S., & Khan, F. A. (1990). Control of root-knot nematodes, *Meloidogyne* spp., on tomato, *Lycopersicon esculentum* Mill., with poultry manure. *International Journal of Pest Management*, 36(4), 332-335.
- Choleva, B., Bileva, T., & Tsvetkov, J. (2007). Organobiological means and methods for control of plant parasitic nematodes as alternative of agrochemicals. *Ecology and Future (Bulgaria)*, 6(4), 43-49.
- Choleva, B., Bileva, T., Tsvetkov, Y., & Barakov, P. (2005). Preliminary study of the green algae *Chlorella* (*Chlorella vulgaris*) for control on the root-knot nematode (*Meloidogyne arenaria*) in tomato plants and ectoparasite *Xiphinema* index in grape seedlings. *Communications in agricultural and applied biological sciences*, 70(4), 915-926.
- Choleva, B., Tsvetkov, Y., & Nedelchev, S. (2004). Non-chemical control of root-knot nematodes (*Meloidogyne* complex) in glasshouse conditions and of *Xiphinema* index in vineyards. In Proceedings of the Fourth International Congress of Nematology, 8-13 June 2002, Tenerife, Spain (pp. 159-164). Brill.
- Collange, B., Navarrete, M., Peyre, G., Mateille, T., & Tchamitchian, M. (2011). Root-knot nematode (*Meloidogyne*) management in vegetable crop production: The challenge of an agronomic system analysis. *Crop protection*, 30(10), 1251-1262.
- Daneel, M., Engelbrecht, E., Fourie, H., & Ahuja, P. (2018). The host status of Brassicaceae to *Meloidogyne* and their effects as cover and biofumigant crops on root-knot nematode populations associated with potato and tomato under South African field conditions. *Crop protection*, 110, 198-206.
- Dar, S. A., Wani, S. H., Mir, S. H., Showkat, A., Dolkar, T., & Dawa, T. (2021). Biopesticides: mode of action, efficacy and scope in pest management. *Journal of Advanced Research in Biochemistry and Pharmacology*, 4(1), 1-8.
- DIRECTIVE 2009/128/EC OF THE EUROPEAN PARLIAMENT AND OF THE COUNCIL of 21 October 2009 establishing a framework for Community action to achieve the sustainable use of pesticides. O.J.E.U., L.309/71, November, 24 2009.
- El-Nagdi, W. E., & Youssef, M. M. A. (2013). Comparative efficacy of garlic clove and castor seed aqueous extracts against the root-knot nematode, *Meloidogyne incognita* infecting tomato plants. *Journal of plant protection research*, 53(3), 285-288.
- Goswami, J., Pandey, R. K., Tewari, J. P., & Goswami, B. K. (2008). Management of root knot nematode on tomato through application of fungal antagonists, *Acremonium strictum* and *Trichoderma harzianum*. *Journal of Environmental Science and Health Part B*, 43(3), 237-240.
- Guimarães, P. M., Brasileiro, A. C. M., Proite, K., de Araújo, A. C. G., Leal-Bertioli, S. C. M., Pic-Taylor, A., ... & Bertioli, D. J. (2010). A study of gene expression in the nematode resistant wild peanut relative, *Arachis stenosperma*, in response to challenge with *Meloidogyne arenaria*. *Tropical Plant Biology*, 3, 183-192.
- Hassan, M. A., Pham, T. H., Shi, H., & Zheng, J., 2013. Nematodes threats to global food security. *Acta Agriculturae Scandinavica, Section B-Soil & Plant Science*, 63(5), 420-425.
- Jindapunnapat, K., Chinnasri, B., & Kwankuae, S. (2013). Biological control of root-knot nematodes (*Meloidogyne enterolobii*) in guava by the fungus *Trichoderma harzianum*. *Journal of Developments in Sustainable Agriculture*, 8(2), 110-118.
- Ladner, D. C., Tchounwou, P. B., & Lawrence, G. W. (2008). Evaluation of the effect of ecologic on root knot nematode, *Meloidogyne incognita*, and tomato plant, *Lycopersicon esculentum*. *International Journal of Environmental Research and Public Health*, 5(2), 104-110.
- Lamovšek, J., Urek, G., & Trdan, S. (2013). Biological control of root-knot nematodes (*Meloidogyne* spp.): microbes against the pests. *Acta Agriculturae Slovenica*, 101(2), 263-275.
- Li, X. Q., Wei, J. Z., Tan, A., & Aroian, R. V. (2007). Resistance to root-knot nematode in tomato roots expressing a nematicidal *Bacillus thuringiensis* crystal protein. *Plant biotechnology journal*, 5(4), 455-464.
- Lizardo, R. C. M., Pinili, M. S., Diaz, M. G. Q., & Cumagun, C. J. R. (2022). Screening for Resistance in Selected Tomato Varieties against the Root-Knot

- Nematode *Meloidogyne incognita* in the Philippines Using a Molecular Marker and Biochemical Analysis. *Plants*, 11(10), 1354.
- Lykogianni, M., Bempelou, E., Karamaouna, F., & Aliferis, K. A. (2021). Do pesticides promote or hinder sustainability in agriculture? The challenge of sustainable use of pesticides in modern agriculture. *Science of the Total Environment*, 795, 148625.
- Markova, D., & Samaliev, H. (2011). Pathogenicity of the root-knot nematode *Meloidogyne hapla* on potato in Bulgaria. *Agrarni Nauki*, 3(7), 71-76.
- Markova, L., Tsolova, E., Kokudev, B., & Bulgaria, T. A. (2014). Influence of the Liquid Root Biostimulator Fertiactyl® GZ on Damaged Tomato Plants Caused by Root-Knot Nematodes *Meloidogyne* spp. *Plant Science (Bulgaria)*, 51(2-3), 97-101.
- Mateeva A., 2004. Not insect enemies. Sofia: PublishSySet-Eco, pp. 238.
- Mesa-Valle, C. M., Garrido-Cardenas, J. A., Cebrian-Carmona, J., Talavera, M., & Manzano-Agugliaro, F., (2020). Global research on plant nematodes. *Agronomy*, 10(8), 1148.
- Mohamedova, M. S. (2009). Efficacy of *Bacillus thuringiensis* alone and in combination with oxamyl against *Meloidogyne arenaria* infecting greenhouse tomato. *Agricultural Science and Technology*, 1(2), 41-44.
- Mohamedova, M., and H. Samaliev (2011). Effect of rhizobacterium *Bacillus subtilis* on the development of root-knot nematode *Meloidogyne arenaria* at difference temperature. *Agricultural Science and Technology*, 3, 378-383.
- Mohamedova, M., Donka, D., Iliyana, V., & Mladen, N. (2016). Effects of rhizobacteria on *Meloidogyne javanica* infection on eggplants. *African Journal of Agricultural Research*, 11(41), 4141-4146.
- Monfort, W. S., Csinos, A. S., Desaegeer, J., Seebold, K., Webster, T. M., & Diaz-Perez, J. C. (2007). Evaluating Brassica species as an alternative control measure for root-knot nematode (*M. incognita*) in Georgia vegetable plasticulture. *Crop Protection*, 26(9), 1359-1368.
- Moreira, F. J. C., Silva, M. C. B., Rodrigues, A. A., & Tavares, M. K. N. (2015). Alternative control of root-knot nematodes (*Meloidogyne javanica* and *M. enterolobii*) using antagonists. *International Journal of Agronomy and Agricultural Research*, 7(2), 121-129.
- Mostafa, M. A., Mahmoud, N. A. B., Anany, A. E. A., & El-Sagheer, A. M. B. (2017). Plant essential oils as eco-friendly management tools for root knot nematode on cucumber plants. *Journal of Zoology Studies*, 4(1), 1-5.
- Motti, R. (2021). The Solanaceae family: Botanical features and diversity. The wild solanums genomes, 1-9. Springer, Cham. Santamaria, & Signore
- Natarajan, N., Cork, A., Boomathi, N., Pandi, R., Velavan, S., & Dhakshnamoorthy, G. (2006). Cold aqueous extracts of African marigold, *Tagetes erecta* for control tomato root knot nematode, *Meloidogyne incognita*. *Crop Protection*, 25(11), 1210-1213.
- Nicol, J. M., Turner, S. J., Coyne, D. L., Nijs, L. D., Hockland, S., & Maafi, Z. T. (2011). Current nematode threats to world agriculture. *Genomics and molecular genetics of plant-nematode interactions*, 21-43. Springer, Dordrecht.
- Oduor-Owino, P. (2003). Control of root-knot nematodes in Kenya with aldicarb and selected antagonistic plants. *Nematologia Mediterranea*, 31(1), 125-127.
- Qiao, K., Liu, X., Wang, H., Xia, X., Ji, X., & Wang, K. (2012). Effect of abamectin on root-knot nematodes and tomato yield. *Pest management science*, 68(6), 853-857.
- Radwan, M. A., Farrag, S. A. A., Abu-Elamayem, M. M., & Ahmed, N. S. (2012). Biological control of the root-knot nematode, *Meloidogyne incognita* on tomato using bioproducts of microbial origin. *Applied Soil Ecology*, 56, 58-62.
- Ravichandra, N. G., 2014. Horticultural nematology (Vol. 2014, New Dehli: Springer India.
- Ravichandra, N.G. (2014). Novel Methods of Nematode Management. In: *Horticultural Nematology*, 412, Springer, New Delhi.
- Saeedzadeh, A., Niasti, F., Baghaei, M. A., Hasanpour, S., & Agahi, K. (2020). Effects of fertilizers on development of root-knot nematode, *Meloidogyne javanica*. *International Journal of Agriculture & Biology*, 23(2), 431-437.
- Salim, H. A., Salman, I. S., Majeed, I. I., & Hussein, H. H. (2016). Evaluation of some plant extracts for their nematocidal properties against root-knot nematode, *Meloidogyne* sp. *Journal of Gene c and Environmental Resources Conservation*, 4(3), 241-244.
- Samaliev, H. (2009a). Races of four species of root-knot nematodes of *Meloidogyne* Goeldi on vegetable crops in glasshouses in Bulgaria. *Rasteniev'dni Nauki*, 46(6), 542-547.
- Samaliev, H. (2009b). Solarization of soil for the control of root-knot nematodes (*Meloidogyne* species) vegetable glasshouses in Southern Bulgaria. *Rasteniev'dni Nauki*, 46(4), 361-365.
- Samaliev, H. Y., Salkova, D. S., Baycheva, O. T., Zinovieva, S. V., & Udalova, Z. V. (2018). Investigations of the root-knot nematodes of the genus *Meloidogyne* (Goeldi, 1887) on the territories of Bulgaria and Russian Federation. *Rossiiskii Parazitologicheskii Zhurnal*, (4), 94-98.
- Samaliev, H., & Baicheva, O. (2010). Distribution of root-knot nematodes (genus *Meloidogyne* Goeldi) on the potatoes in the Plovdiv, Troyan and Samokov regions in Bulgaria. In The Materials of the International Conference (November 30-December 3, 2010, Moscow) (pp. 458-465).
- Samaliev, H., & Kalinova, S. (2013). Host suitability of twelve common weeds to *Pratylenchus penetrans* and *Meloidogyne hapla* in potato fields of Bulgaria. *Bulgarian Journal of Agricultural Science*, 19(2), 202-208.
- Samaliev, H., Andreoglou, F., Elawad, S., Hague, N., & Gowen, S. (2000). The nematocidal effects of the bacteria *Pseudomonas oryziphobans* and *Xenorhabdus nematophilus* on the root-knot nematode *Meloidogyne javanica*. *Nematology*, 2(5), 507-514.



- Samaliev, H., Stoyanov D., (2008). Parasitic nematodes on crop plants and the fight against them. Agricultural University Academic Press, 350.
- Santamaria, P., & Signore, A., 2021. How has the consistency of the Common catalogue of varieties of vegetable species changed in the last ten years? *Scientia Horticulturae*, 277, 109805.
- Shaikh, S. S., & Sayyed, R. Z., 2015. Role of plant growth-promoting rhizobacteria and their formulation in biocontrol of plant diseases. In *Plant microbes symbiosis: Applied facets* (pp. 337-351). Springer, New Delhi.
- Sidhu, H. S. (2018). Potential of plant growth-promoting rhizobacteria in the management of nematodes: a review. *Journal of Entomology and Zoology studies*, 6(3), 1536-1545.
- Singh, A., Sharma, B., Kumari, A., Kumar, R., & Pathak, D. V. (2019). Management of root-knot nematode in different crops using microorganisms. In *Plant biotic interactions* (pp. 85-99). Springer, Cham.
- Singh, S., Singh, B., & Singh, A. P. (2015). Nematodes: a threat to sustainability of agriculture. *Procedia Environmental Sciences*, 29, 215-216.
- Sosnowska, D. (2007). Effect of Host Plant and Nematophagous Fungi on Population of the Peanut Root-Knot Nematode (*Meloidogyne arenaria* (Neal) Chitwood. *Rasteniev'dni Nauki* 44(3), 240.
- Stirling, G. R., & Stirling, A. M. (2003). The potential of Brassica green manure crops for controlling root-knot nematode (*Meloidogyne javanica*) on horticultural crops in a subtropical environment. *Australian Journal of Experimental Agriculture*, 43(6), 623-630.
- Stoyanov, D. (1980). Identification of the host plants of gall nematodes from the genus *Meloidogyne* Goeldi, 1887 in Bulgaria. *Rasteniev'dni Nauki*, 17(3), 65-78.
- Taniwiryono, D., Berg, H., Riksen, J. A. G., Rietjens, I. M. C. M., Djiwantia, S. R., Kammenga, J. E., & Murk, A. J. (2009). Nematicidal activity of plant extracts against the root-knot nematode, *Meloidogyne incognita*. *The Open Natural Products Journal*, 2(1).
- Taye, W., Sakhuja, P. K., & Tefera, T. (2012). Evaluation of plant extracts on infestation of root-knot nematode on tomato (*Lycopersicon esculentum* Mill). *E3 Journal of Agricultural Research and Development*, 2(3), 086-091.
- Trifonova ZI, J. Karadjova, Georgieva Tr., (2009). Fungal parasites of the root-knot nematodes *Meloidogyne* spp. in southern Bulgaria. *Estonian Journal of Ecology*, 58, 1, 47-52.
- Trifonova, Z. (2014). Studies on the efficacy of some fungi and biopreparations for control of *Meloidogyne incognita*. *Plant Science (Bulgaria)*. 51, 4-5, 12-15.
- Trifonova, Z., & Atanasov, A. (2009). Investigation of the nematicidal effects of some plant extracts on the mortality and the hatching of *Meloidogyne incognita* and *Globodera rostochiensis*. *Rasteniev'dni Nauki*, 46(6), 548-553.
- Trifonova, Z., (2012). Control of *Globodera rostochiensis* and *Meloidogyne incognita* with Some Plant Oils by Bare-Root Dip Treatment. *Bulgarian Journal of Ecological Science (Bulgaria)*. 11, 3, 35-39.
- Trifonova, Z., & Vachev, T. (2010). Efficacy of *Trichoderma* species on invasion and development of root-knot nematode *Meloidogyne incognita* in tomato agroecosystem. *Journal of Balkan Ecology*, 13(4), 379-384.
- Trifonova, Z., & Voulkova, Z. (2008). Response of F1 hybrids and their parental forms of *Lycopersicon* genus to *Meloidogyne incognita* and *M. hapla*. *Zaštita bilja*, 59(1-4), 93-99.
- Trifonova, Z., & Vulkova, Z. (2007). Resistance of F1 hybrids of *Lycopersicon* genus to populations of *Meloidogyne* species. *Bulgarian Journal of Agricultural Science*, 13(5), 535.
- Trifonova, Z., Karadjova, J., & Georgieva, T. (2009). Fungal parasites of the root-knot nematodes *Meloidogyne* spp. in Southern Bulgaria., *Estonian Journal of Ecology*, 58, 1, 47-52
- Tringovska, I., Yankova, V., Markova, D., & Mihov, M. (2015). Effect of companion plants on tomato greenhouse production. *Scientia Horticulturae*, 186, 31-37.
- Verma, P. P., Shelake, R. M., Das, S., Sharma, P., & Kim, J. Y. (2019). Plant growth-promoting rhizobacteria (PGPR) and fungi (PGPF): potential biological control agents of diseases and pests. *Microbial Interventions in Agriculture and Environment: Volume 1: Research Trends, Priorities and Prospects*, 281-311.
- Volpiano, C. G., Lisboa, B. B., Granada, C. E., José, J. F. B. S., de Oliveira, A. M. R., Beneduzi, A., ... & Vargas, L. K. (2019). Rhizobia for biological control of plant diseases. *Microbiome in plant health and disease: challenges and opportunities*, 315-336.
- Voulkova, Z., & Trifonova, Z. (2009). Response of wild species of *Lycopersicon* genus to *Meloidogyne* spp. and *Globodera rostochiensis* Woll. *Journal of Balkan Ecology*, 12(3), 257-262.
- Yankova, V., Ganeva, D., & Loginova, E. (2006). Study of tomato breeding lines response to root knot nematode infestation *Meloidogyne arenaria* Neal. *Plant Science*.43, 4,356-360.
- Yankova, V., Markova, D., naidenov, M., & Arnaudov, B. (2014). Management of root-knot nematodes (*Meloidogyne* spp.) in greenhouse cucumbers using microbial products. *Türk Tarım ve Doğa Bilimleri Dergisi*, 1(Özel Sayı-2), 1569-1573.
- Yasmin, L., Rashid, M. H., Uddin, M. N., Hossain, M. S., Hossain, M. E., & Ahmed, M. U. (2003). Use of neem extract in controlling root-knot nematode (*Meloidogyne javanica*) of sweet-gourd. *Pakistan Journal of Plant Pathology (Pakistan)*, 2(3),161-168.
- Yücel, S., Özarslandan, A., Colak, A., Ay, T., & Can, C. (2007). Effect of solarization and fumigant applications on soilborne pathogens and root-knot nematodes in greenhouse-grown tomato in Turkey. *Phytoparasitica*, 35(5), 450-456.
- Zakaria, H. M., Kassab, A. S., Shamseldean, M. M., Oraby, M. M., & El-Mourshedy, M. M. F. (2013). Controlling the root-knot nematode, *Meloidogyne incognita* in cucumber plants using some soil bioagents and some amendments under simulated field conditions. *Annals of Agricultural Sciences*, 58(1), 77-82.
- Zavaleta-Mejia, E., & VanGundy, S. D. (1982). Effects of rhizobacteria on *Meloidogyne* infection. *Journal Nematology*. 14(1),475A-475B.