

## MOLECULAR SCREENING OF APPLE CULTIVARS FOR TWO SCAB RESISTANCE GENES IN ROMANIA

Adina Floricica IANCU<sup>1,2</sup>, Mădălina MILITARU<sup>2</sup>, Monica STURZEANU<sup>2</sup>,  
Eugenia MAREȘI<sup>2</sup>

<sup>1</sup>Doctoral School of Plant and Animal Resources Engineering, Faculty of Horticulture,  
University of Craiova, 13 A. I. Cuza Street, 200585 Craiova, Romania

<sup>2</sup>Research Institute for Fruit Growing Pitești-Mărăcineni, 402 Mărului Street,  
117450, Mărăcineni, Argeș, Romania

Corresponding author email: adinafloricica@yahoo.com

### Abstract

*Apple scab, caused by Venturia inaequalis, is considered the most damage disease of the apple growing in Romania. Pyramiding multiple sources of quantitative resistance could be the best way to control the attack of this fungi. A set of forty apple cultivars from the apple gene bank in Pitesti, Romania was evaluated for the presence of two scab resistance genes: Rvi8 and Rvi11. Using marker OPB18, it was made the difference between 2 genes, Rvi2 and Rvi8, located in the same locus (Vr gene). For identification of Rvi11, the SCAR marker, K08, was used and seventeen cultivars amplified the 743 bp fragment that is linked with Rvi11 gene. The results on resistance of apple Romanian cultivars towards economically important disease is necessary for future breeding work and for establishing of new commercial orchards.*

**Key words:** breeding; genotypes; Rvi2; Rvi8; Rvi11.

### INTRODUCTION

SCAR markers were first developed and initially applied to study on resistance genes for downy mildew in lettuce by Paran and Michelmore (1993). A SCAR marker is a technique mediated by PCR, which identifies the genomic DNA fragment at a single locus, using a pair of specific oligonucleotides, of 15-30 bp, projected from nucleotide sequences derived from dominant polymorphic RAPD fragments.

To highlight some apple resistance genes, the most used markers associated were: AL-07, AM19, Vfc, OPL19, OPB12, OPB18, AD13, OPB12, K08, T06, Z13, 41A24T7, ARGH17 and for resistance to powdery mildew, the most used markers were: OPU02, EM M01, AT20, EMDM01, EM 02. Most of them (Al07, AM19, Vfc, OPL19, AD13, OPB12, AT20, EM M01) were used in the studies initiated at the Research Institute for Fruit Growing Pitești-Mărăcineni (Militaru et al., 2020; Iancu et al., 2022; Militaru et al., 2022; Iancu et al., 2023). OPL19<sub>433</sub> was developed by cloning the 550 bp fragment from the RAPD marker OPL19<sub>550</sub>, located near the Rvi2 gene, at a distance of 2.5

cM (Gardiner et al. 1999a; Bus et al., 1999). The SCAR marker was also mapped to 1 cM (Bus et al., 2005b) close to Rvi2 (Vh2) in differential host 2 ('TSR34T15'), but also close to the Rvi8 gene (Vh8) in differential host 8 (*M. sieversii* W193B), being considered very useful for identifying varieties carrying the Vr gene (Rvi2 or Rvi8).

To differentiate the carrier varieties of one of the genes Rvi2 or Rvi8, Bus et al. (2005a), developed the OPB18 SCAR marker, which amplified in a population 'Royal Gala × *M. sieversii* W193B' only those hybrids carrying of the Rvi8 gene.

The Rvi11 gene was identified in *Malus baccata* by Dayton and Williams in 1968 and was mapped to LG-2 by Gessles et al. in 2006. Gyax et al. (2004) showed the presence of the Rvi11 gene (old name Vbj) using the SCAR markers K08<sub>743</sub>, Z13<sub>773</sub>, T06<sub>410</sub> which were projected from the decamer predecessors OPK08<sub>848</sub>, OPZ13<sub>869</sub> and OPT06<sub>801</sub>.

Galli et al. (2010) revealed three transcribed putative resistance gene analogues (Resistance Gene Analogs) for resistance to *Venturia inaequalis* (Vr2-A, Vr2-B and Vr2-C) after isolation of Bacterial Artificial Chromosome

(BAC) spanning the *Rvi15* (*Vr2*) region and it sequenced, and Schouten et al. (2014) concluded that Vr2-C is the *Rvi15* (*Vr2*) gene that provides the resistance. Markers 41A24T7 and 43M10RP have cosegregated on the TIR-NBS-LRR domain of the *Vr2-B*. The Vr2-C region was flanked by the markers: 77G20RP (SSR), 21K14T7 (SNP), 8K11RP (SCAR) and GmTNL1 (SNP). Two other flanking markers ARGH17 (CAPS) and 48K16T7 (SCAR) cover the region of the *Vr2-A* gene. ARGH17 and GmTNL1 were mapped closest to both regions of the resistance locus, *Vr2-A* (0.3 cM) and *Vr2-C* (0.2 cM), respectively (Galli et al., 2010).

## MATERIALS AND METHODS

### Biological material studied

Forty apple cultivars ('Alex', 'Aura', 'Auriu de Cluj', 'Bistrițean', 'Cezar', 'Ciprian', 'Colmar', 'Colonade', 'Dacian', 'Dany', 'Delicios de Voinești', 'Discoprim', 'Doina', 'Estival', 'Frumos de Voineti', 'Generos', 'Inedit', 'Iris', 'Irisem', 'Ionaprim', 'Luca', 'Nicol', 'Pomona', 'Precoce de Ardeal', 'Productiv de Cluj', 'Real', 'Rebra', 'Redix', 'Revidar', 'Remar', 'Remus', 'Romus 3', 'Romus 4', 'Romus 5', 'Rustic', 'Salva', 'Starkprim', 'Voinicel', 'Voinea', 'Valery'), which belong to the gene bank located at Research Institute for Fruit Growing Pitesti, Romania were used in the study.

Three leaf samples from each three trees per cultivar were collected. For the identification of the scab resistance genes, *Rvi8* and *Rvi11*, the following SCAR markers were applied: OPB18 and K08, respectively (Table 1).

'*Malus baccata jackii*' was used as a positive control for the K08 marker associated with the scab resistance gene *Rvi11*.

### Chemical material

The DNA was extracted according to the recommended working method of the "Isolate II Plant DNA Kit" protocol. For the migration of the amplified fragments, a 3% agarose gel was prepared in TBE 1X buffer and subsequently stained with "RedSafe Staining" Nucleic Acid. The amplification of the reactions was performed using the "2x PCR BIO Taq Mix Red" kit from Biosystems.

### PCR amplification

PCR amplification was performed using the "FastGene" analyzer. The PCR reaction was performed separately for each of the primer pairs, in a reaction volume of 15 µl, of which: 12 µl "2x PCR BIO Taq Mix Red"; 0.1 µl from every marker and 3 µl DNA (75 ng/µl) for both markers. The reaction conditions were: initial denaturation at 94°C for 3 min; 40 cycles of 1 min at 94°C, 1 min at T<sub>m</sub> and 2 min at 72°C; final extension of 10 min at 72°C (65°C for K08, 55°C for OPB18).

Table 1. SCAR molecular markers for *Rvi8* and *Rvi11* genes

Gene	Name primer	Primer sequences	Fragment size (bp)	References
<i>Rvi8</i> ( <i>Vh8</i> )	OPB18	F: CCACAGCAGTCATTGGGA R: CCACAGCAGTGCATAAAC	628; 799	Bus et al, 2005a
<i>Rvi11</i> ( <i>Vbj</i> )	K08	F: GAACACTGGGCAAAGGAAAC R: TAAAAGCCACGTTCTCTCGC	743; 900	Gzgax et al., 2004

### Evaluation of results

The fragments amplified following the PCR reaction were loaded in a volume of 10 µl for each sample, in the agarose gel and read with a high-quality "Uvitec Cambridge Essential" imaging system using UVITec1D analysis software. The duration of the sample migration was 4 hours at a voltage of 50 volts for gels with a concentration of 3% and a horizontal electrophoresis system "Wide Midi Horizontal Electrophoresis System" from Cleaver Scientific was performed.

### Statistical analysis

Statistical analysis has been used to order varieties with polygenic characteristics into groups and subgroups. This grouping was made with the GeneAlex v software. 6.51b2. The genotype-phenotype correlation was calculated with the Pearson correlation coefficient, using Minitab18 software.

The statistical analysis of allelic polymorphism, taking into account only the dominant allele of the genes of interest, was expressed using the PIC index (Content of polymorphic

information), which takes into account the allelic frequency, being calculated using mathematical expression:  $2f(1-f)$ . Two statistical indices were used to quantify genetic diversity: the Shannon index and the Simpson index. The Shannon index was calculated with the mathematical formula:  $-\sum_{i=1}^n \frac{n_i}{N} * \ln n_i/N$  and the Simpson index with the formula:  $(\frac{\sum_{i=1}^n n_i * (n_i - 1)}{N * (N - 1)})$ , where:  $n$  represents the allele at the monolocus level, and  $N$  - the total number of alleles (Shannon et al., 1948; Simpson, 1960).

## RESULTS AND DISCUSSIONS

**Rvi8.** The SCAR markers OPB18 and OPB19 are indicators for *Rvi8* and *Rvi2* genes. Iancu et al. (2022, 2023) using marker OPL19 reported an allele size of 433 bp for *Vr* gene (*Rvi2* or

*Rvi8*) for the for twenty cultivars: 'Alex', 'Bistrițean', 'Cezar', 'Ciprian', 'Dany', 'Discoprim', 'Delicios de Voinești', 'Estival', 'Jonaprim', 'Luca', 'Pomona', 'Romus 3', 'Romus 4', 'Romus 5', 'Redix', 'Remar', 'Salva', 'Starkprim', 'Voinecel', 'Voinea'.

The segregation of the OPB18 marker with the gene locus made the difference between the *Rvi8* and *Rvi2* genes, producing the amplification of the 799 bp fragment associated with the dominant allele of the *Rvi8* gene, only. This gene did not segregate into the apple cultivars studied, and, in order to confirm our results, other two cultivars 'Verzișoare' and 'Pionier' were, supplementary, introduced, as control. The OPL18 marker amplified the 799 bp fragment corresponding to the dominant allele of the *Rvi8* gene for both control cultivars (Figure 1).

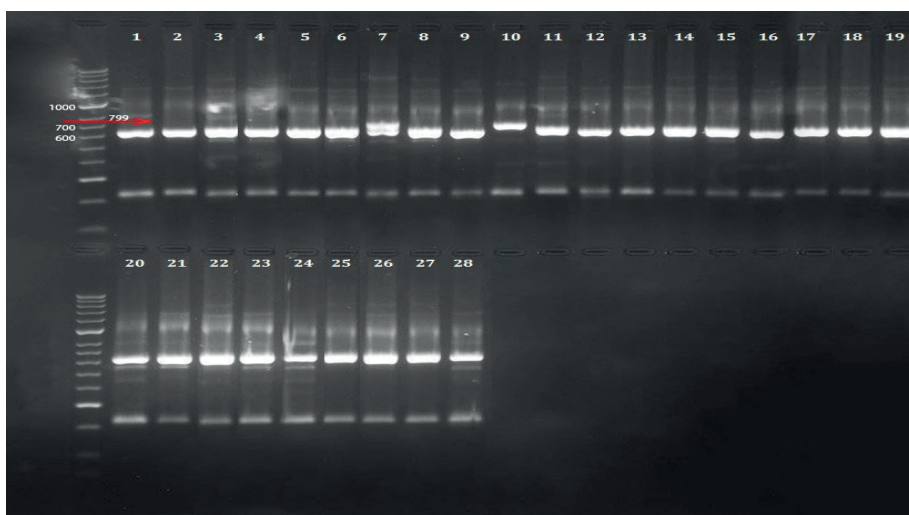


Figure 1. Electrophoretic profile performed with the OPB18 marker for genotypes in which specific amplifications of the *Vr* locus were obtained with the OPL19 marker:

1. 'Romus 2', 2. 'Romus 3', 3. 'Romus 4', 4. 'Romus 5', 5. 'Florina', 6. 'Prima', 7. 'Pionier', 8. 'Starkrimson', 9. 'Crețesc', 10. 'Verzișoare', 11. 'Parmen d'or', 12. 'Estival', 13. 'Bistrițean', 14. 'Dany', 15. 'Luca', 16. 'Ciprian', 17. 'Salva', 18. 'Jonaprim', 19. 'Starkprim', 20. 'Delicios de Voinești', 21. 'Redix', 22. 'Alex', 23. 'Voinecel', 24. 'Voinea', 25. 'Discoprim', 26. 'Cezar', 27. 'Pomona', 28. 'Remar'

**Rvi11.** The codominant marker K08 was segregated into 19 cultivars ('Alex', 'Bistrițean', 'Colonade', 'Ciprian', 'Colmar', 'Dany', 'Dacian', 'Estival', 'Generos', 'Irisem', 'Nicol', 'Pomona', 'Precoce de Ardeal', 'Productiv de Cluj', 'Rebra', 'Romus 3', 'Remus', 'Redix', 'Remar') and '*Malus baccata jacksonii*', as positive control. This

marker made the difference between heterozygous and homozygous genotypes by amplifying the 743 bp and 900 bp, respectively: 11 were heterozygous with both dominant and recessive alleles, and 8 were homozygous with dominant alleles (Figure 2).

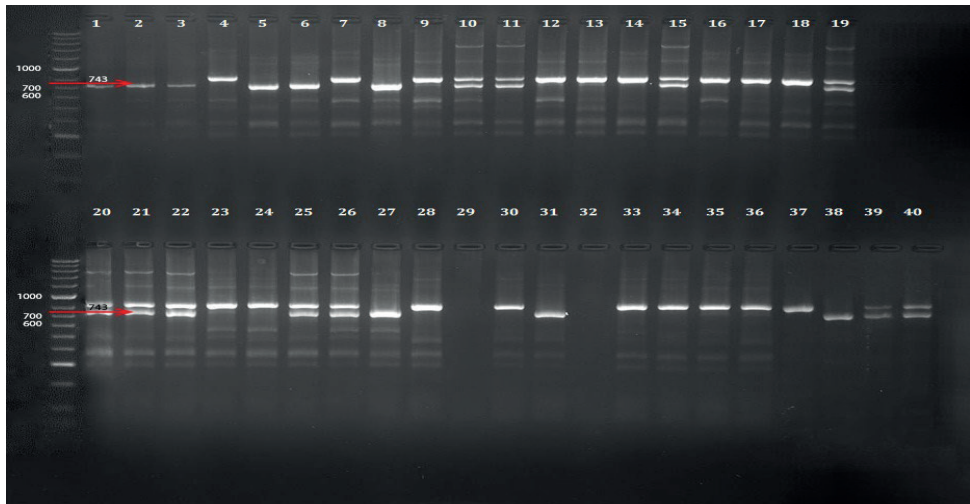


Figure 2. Electrophoretic profile released with the K08 marker:

1. 'Estival', 2. 'Rebra', 3. 'Bistrițean', 4. 'Aura', 5. 'Romus 3', 6. 'Dany', 7. 'Romus 4', 8. 'Productiv de Cluj', 9. 'Luca', 10. 'Ciprian', 11. 'Irisem', 12. 'Slava', 13. 'Ionaprim', 14. 'Rustic', 15. 'Precoce de Ardeal', 16. 'Iris', 17. 'Starkprim', 18. 'Auriu de Cluj', 19. 'Generos', 20. 'Colonade', 21. 'Nicol', 22. 'Colmar', 23. 'Delicios de Voinești', 24. 'Romus 5', 25. 'Remus', 26. 'Redix', 27. 'Alex', 28. 'Doina', 29. 'Voincei', 30. 'Inedit', 31. 'Dacian', 32. 'Voinea', 33. 'Valery', 34. 'Real', 35. 'Discoprim', 36. 'Cezar', 37. 'Frumos de Voinești', 38. 'Pomona', 39. 'Revidar', 40. 'Remar'

By cumulating the present with previous results, we obtained a scab resistance genetic profile for all forty apple cultivars (Table 2). The data showed different scab gene accumulations, such as: tetragenic (*Rvi2+Rvi4+Rvi6+Rvi11*), trygenic

(*Rvi2+Rvi4+Rvi6; Rvi2+Rvi4+Rvi11; Rvi2+Rvi6+Rvi11*), digenic (*Rvi2+Rvi6; Rvi6+Rvi11; Rvi5+Rvi11*), monogenic (*Rvi2; Rvi6; Rvi11*). For two cultivars ('Frumos de Voinești' and 'Auriu de Cluj') which did not show any scab resistance genes.

Table 2. Scab resistance genetic profile

Cultivar	<i>Rvi8</i> <sup>a</sup>	<i>Rvi11</i> <sup>a</sup>	<i>Rvi2</i> <sup>b</sup>	Genetic profile <sup>a,b</sup>
	OPB18	K08	OPL19	
Estival	.-	+	+	<i>Rvi2+Rvi4 +Rvi11</i>
Rebra	-	+	-	<i>Rvi6+Rvi11</i>
Bistrițean	.-	+	+	<i>Rvi2+Rvi6 +Rvi11</i>
Aura	-	-	-	<i>Rvi6</i>
Romus 3	.	+	+	<i>Rvi2+Rvi6+Rvi4+Rvi11</i>
Dany	.	+	+	<i>Rvi2+Rvi6 +Rvi11</i>
Romus 5	.	-	+	<i>Rvi2+Rvi6+Rvi4</i>
Productiv de Cluj	-	+	-	<i>Rvi11</i>
Luca	-	-	+	<i>Rvi2+Rvi6</i>
Ciprian	-	+	+	<i>Rvi2+Rvi6 +Rvi11</i>
Irisem	-	+	-	<i>Rvi5+ Rvi11</i>
Salva	-	-	+	<i>Rvi2+Rvi6</i>
Ionaprim	-	-	+	<i>Rvi2+Rvi6</i>
Rustic	-	-	-	<i>Rvi6</i>
Precoce de Ardeal	-	+	-	<i>Rvi11</i>
Iris	-	-	-	<i>Rvi6</i>
Starkprim	-	-	+	<i>Rvi2+Rvi6</i>
Auriu de Cluj	-	-	-	-
Generos	-	+	-	<i>Rvi5+ Rvi11</i>
Colonade	-	+	-	<i>Rvi6+Rvi11</i>
Nicol	-	+	-	<i>Rvi5+ Rvi11</i>
Colmar	-	+	-	<i>Rvi6+Rvi11</i>
Delicios de Voinești	-	-	+	<i>Rvi2</i>
Romus 4	.	-	+	<i>Rvi2+Rvi6</i>

Remus	-	+	-	<i>Rvi11</i>
Redix	-	+	+	<i>Rvi2+Rvi6+Rvi4+Rvi11</i>
Alex	-	+	+	<i>Rvi2+Rvi6 +Rvi11</i>
Doina	-	-	-	<i>Rvi6</i>
Voinicel	-	-	+	<i>Rvi2+Rvi6+Rvi4</i>
Inedit	-	-	-	<i>Rvi6</i>
Dacian	-	+	-	<i>Rvi6+Rvi11</i>
Voinea	+	-	-	<i>Rvi2+Rvi6</i>
Valery	-	-	-	<i>Rvi6</i>
Real	-	-	-	<i>Rvi6</i>
Discoprim	-	-	+	<i>Rvi2+Rvi6+Rvi4</i>
Cezar	-	-	+	<i>Rvi2+Rvi6+Rvi4</i>
Frumos de Voinești	-	-	-	-
Pomona	-	+	+	<i>Rvi2+Rvi6+Rvi4+Rvi11</i>
Revidar	-	-	-	<i>Rvi6+Rvi11</i>
Remar	-	+	+	<i>Rvi2+Rvi6+Rvi4+Rvi11</i>

<sup>a</sup>Molecular screening conducted in this study

<sup>b</sup>Iancu et al., 2023

In figure 3, using the statistical method PCoA analysis (standard covariance) with the help GeneAlex v software. 6.51b2, it can observe

the distribution of cultivars which carried dominant alleles for scab resistance in groups and subgroups.

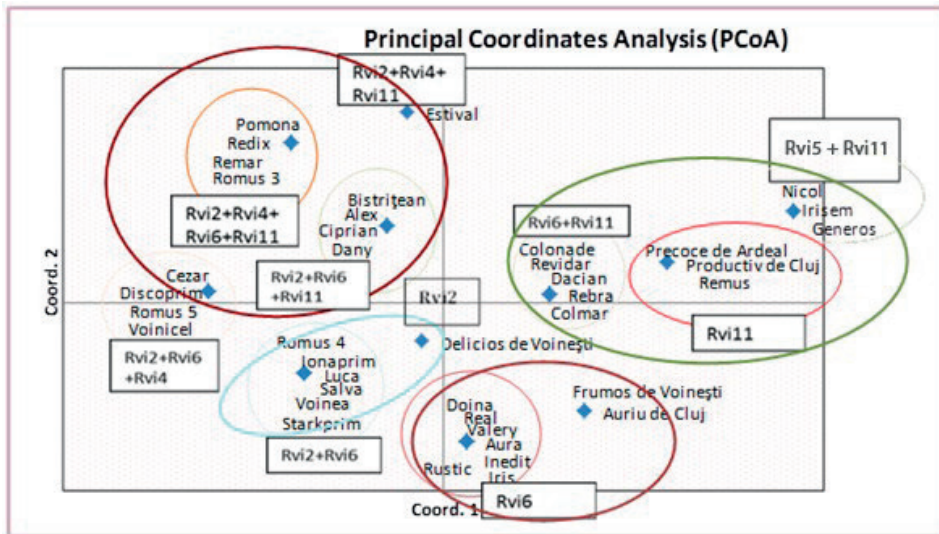


Figure 3. Distribution of cultivars based on R genes screening

Correlating phenotypic data presenting by Militaru et al. (2022) with our genotype results, we conclude that five of the monogenic varieties ('Aura', 'Inedit', 'Iris', 'Valery', 'Real') have a scab resistant response, while the varieties 'Doina' and 'Rustic', with the same monogenic characteristic (*Rvi6* gene), show a moderate response.

Some varieties with trigenic characteristics from the subgroup "*Rvi2+Rvi6+Rvi11*" ('Bistrițean' and 'Ciprian') are scab resistant, while 'Dany' and 'Alex' varieties, for same

subgroup, manifest a phenotypic response of moderate resistance.

'Colmar', 'Colonade', 'Revidar' and 'Dacian' of the subgroup "*Rvi6+Rvi11*" are resistant to apple scab, while the variety 'Rebra' of the same subgroup shows a "moderate resistance". Also, varieties with digenic characteristics ('Ionaprim', 'Starkprim', 'Luca', 'Voinea' and 'Romus 4') from the subgroup "*Rvi2+Rvi6*" are scab resistant, instead, the 'Salva' variety, from the same subgroup, shows a phenotypic response as "moderate resistance".



The varieties with monogenic characteristics 'Precoce de Ardeal' and 'Productiv de Cluj' (*Rvi11*) have a moderate resistant to apple scab, while 'Remus' (*Rvi11*) and 'Delicios de Voinești' (*Rvi2*) are scab susceptible.

The disease susceptibility response positions the 'Redix' cultivar in the category of plants known as GPI (Genotype-Phenotype Incongruence) as the genotyping result is unexpected, this having a polygenic character: *Rvi2+Rvi4+Rvi6+Rvi11*. The gene combination "*Rvi2+Rvi11*" is not enough to produce an

immune response to the attack of the pathogen, *Venturia inaequalis*. So, the varieties 'Nicol', 'Generos' and 'Irisem' are susceptible to this disease. Using as statistical method the correlation, Pearson coefficient correlated with Minitab18 software, the combination of genes "*Rvi2+Rvi4+Rvi11*" is in perfect correlation with the phenotypic expression manifested by 'Estival' cultivar.

Significant results were also obtained for the combinations of genes "*Rvi2+Rvi6+Rvi11*" and "*Rvi2+Rvi4+Rvi6+Rvi11*" (Figure 4).

	C2	C3	C4	C5	C6	C7	C8	C9	C10	C11	C12
C13	0.031	-0.093	-0.217	0.450	0.015	-0.221	-0.190	-0.202	0.372	0.238	0.277
	<b>0.850</b>	<b>0.568</b>	0.179	0.004	<b>0.927</b>	0.170	0.241	0.212	0.018	0.139	0.083

Figure 4. Pearson correlation between genotypes and phenotypes:

C2 (*Rvi2+Rvi4+Rvi6+Rvi11*: 'Pomona', 'Redix', 'Remar', 'Romus 3'); C3 (*Rvi2+Rvi6+Rvi11*: 'Alex', 'Bistrițean', 'Ciprian', 'Dany'); C4 (*Rvi2+Rvi4+Rvi6*: 'Cezar', 'Discoprim', 'Romus 5', 'Voinicel'); C5 (*Rvi5+Rvi11*: 'Generos', 'Irisem', 'Nicol'); C6 (*Rvi2+Rvi4+Rvi11*: 'Estival'); C7 (*Rvi2+Rvi6*: 'Ionaprim', 'Luca', 'Romus 4', 'Salva', 'Starkprim', 'Voinea'); C8 (*Rvi6+Rvi11*: 'Colonade', 'Colmar', 'Dacian', 'Rebra', 'Revidar'); C9 (*Rvi6*: 'Aura', 'Doina', 'Inedit', 'Iris', 'Real', 'Rustic', 'Valery'); C10 (*Rvi2*: 'Delicios de Voinești'); C11 (*Rvi11*: 'Precoce de Ardeal', 'Productiv de Cluj', 'Remus'); C12 (-); C13 (phenotypic expression)

Genetic diversity and allelic polymorphism were calculated only for the dominant allele of interest genes. The results showed moderately informative polymorphism and low genetic

diversity (Table 3). The small value for genetic diversity is explained by the fact that the same gene has been highlighted in several varieties.

Table 3. Statistical analysis of allelic polymorphism and genetic diversity

PIC value>0.25 (moderately informative)	Shannon Index H > 3 (high genetic diversity)	Simson Index (D) D=0 (infinite diversity) D=1 (lack of diversity)	Simpson diversity index (1-D) value € [0.1]
0.333542	1.294129	0.2494129	0.759944

## CONCLUSIONS

In Romania, the major problem of apple production is scab caused by fungal pathogen, *Venturia inaequalis*. In order to solve this problem, the most Romanian breeding programs included releasing of new cultivars with scab resistance, as major objective. So, the present data complete the information about using of Romanian bred cultivars as a source of diverse functional alleles involved in quantitative resistance against *V. inaequalis* strains. Using K08 marker, it has been established that 19 apple cultivars carriers of *Rvi11* gene and, using OPB18 marker, is not

obtained amplified for *Rvi8*. Genetic diversity of the dominant characteristic for the gene of interest is reduced and allelic polymorphism has a moderately informative value. Segregation of the genes of interest into descendants allowed for validation of the identity of the genitors.

## ACKNOWLEDGEMENTS

The authors express a sincere appreciation for the financial support supported by the ADER project 6.1.4., financed by the Ministry of Agriculture and Rural Development of Romania.

## REFERENCES

- Bus V, Ranatunga C, Gardiner S, Bassett H, Rikkerink E (1999). Marker assisted selection for pest and disease resistance in the New Zealand apple breeding programme. *Acta Hort.* 538: 541–54.
- Bus VGM, Laurens FND, van de Weg WE, Rusholme RL, Rikkerink EHA, Gardiner SE (2005a). The *Vh8* locus of a new gene-for-gene interaction between *Venturia inaequalis* and the wild apple *Malus sieversii* is closely linked to the *Vh2* locus in *Malus pumila* R12740-7A. *New Phytologist*, 166(3): 1035-1049.
- Bus VGM, Rikkerink EHA, van de Weg WE, Rusholme RL, Gardiner SE, Bassett HCM (2005b). The *Vh2* and *Vh4* scab resistance genes in two differential hosts derived from Russian apple R12740-7A map to the same linkage group of apple. *Molecular Breeding*, 15(1): 103-116.
- Choupannejad R, Sharifnabi B, Bahar M, Talebi M (2018). Searching for resistance genes to *Venturia inaequalis* in wild and domestic apples in Iran. *Sci. Hort.* 232: 107-111.
- Dayton DF, Williams EB (1968). Independent genes in *Malus* for resistance to *Venturia inaequalis*, *Prom Am Soc Hort Sci* 92: 89-94
- Galli P, Patocchi A, Brogini G, Gessler C (2010). The *Rvi15 (Vr2)* apple scab resistance locus contains three TIR-NBS-LRR genes. *Molecular Plant-Microbe Interactions*, 23(5): 608-617.
- Gardiner SE, Bus V, Bassett H, Goodman M, Greer L, Ranatunga C, Rikkerink E, Foster R (1999a). Identification of molecular markers linked to major resistances to apple scab, powdery mildew and woolly aphid in apple. In: *Plant and Animal Genome VII Conference*, San Diego, CA, USA, 185.
- Gardiner S, Murdoch J, Meech S, Rusholme R, Rikkerink E, Gleave A, Crowhurst R, Ross G, Bus V, Warrington I, 2003. Candidate resistance genes from an EST database prove a rich source of markers for major genes conferring resistance to important apple pests and diseases. *Acta Hort.* 622: 141-151.
- Gessler C, Patocchi A, Sansavini S, Tartarini S, Gianfranceschi L (2006). *Venturia inaequalis* resistance in apple, *Critical Reviews in Plant Sciences* 25: 473-503
- Gygax M, Gianfranceschi L, Liebhard R, Kellerhals M, Gessler C, Patocchi A (2004). Molecular markers linked to the apple scab resistance gene *Vbj* derived from *Malus baccata jackii*. *Theoretical and Applied Genetics*. 109: 1702-1709.
- Iancu A, Cosmulescu S (2022). The molecular screening of some foreign and Romanian varieties used in breeding programs in Romania. *Fruit Growing Research*, XXXVIII: 76-83.
- Iancu A, Militaru M, Sturzeanu M (2023). Molecular characterization of Romanian apple cultivars for the identification of scab and powdery mildew resistance genes. *Acta Horticulturae* 1384 (52).
- Iancu A, Militaru M, Cosmulescu S (2023). Results regarding concentration, purity, integrity of DNA and optimization of SCAR-SSR multiplex reactions for some Romanian apple varieties. *RIFG Pitesti. Fruit Growing Research*, XXXIX: 62-74.
- Liebhard R, Gianfranceschi L, Koller B, Ryder CD, Tarchini R, van de Weg E, Gessler C (2002): Development and characterization of 140 new microsatellite in apple (*Malus domestica Borkh.*). *Mol. Breeding*. 10:217-241.
- Luo F, Sandefur P, Evans, K, and Peace, C (2019). A DNA test for routinely predicting mildew resistance in descendants of crabapple ‘White Angel’. *Molecular Breeding*. 39, 33. DOI: 10.1007/s11032-019-0933-3
- Militaru M, Sturzeanu M, Iancu A (2020). Molecular screening of some Romanian apple cultivars for scab resistance genes. *Fruit Growing Research*, XXXVI: 5-11.
- Militaru M, Călinescu M, Mareși E, Iancu A, Young-un S, Yong-seub S (2022). Evaluation of scab and powdery mildew resistance of apple germplasm collected at RIFG Pitesti. *Fruit Growing Research*, XXXVIII: 26-31.
- Paran I and Michelmore RW (1993). Development of reliable PCR-based markers linked to downy mildew resistance genes in lettuce. *Theor. Appl. Genet.* 85: 985-993.
- Peil A, Howard NP, Bühlmann-Schütz S, Hiller I, Schouten H, Flachowsky H, Patocchi A. (2023). *Rvi4* and *Rvi15* are the same apple scab resistance genes. *Mol Breed*; 43(10):74. doi: 10.1007/s11032-023-01421-0. PMID: 37830083; PMCID: PMC10564682.
- Schouten HJ, Brinkhuis J, van der Burgh A, Schaart JG, Groenwold R, Brogini GA, Gessler C, (2014). Cloning and functional characterization of the *Rvi15 (Vr2)* gene for apple scab resistance. *Tree Genet Genomes*, 10: 251-260.
- Shannon CE, Weaver W. (1948). *A Mathematical Theory of Communication*. *Bell System Technical Journal*, vol. 27: 379–423, 623–656.
- Simpson GG. (1960). Notes on the measurement of faunal resemblance. *American Journal of Science*, vol. 258A: 300-311.